

Extraction of Eco-Friendly Natural Dyes from *Tradescantia pallida Purpurea* and *Cynomorium coccineum* Growing Naturally in Tunisia

Mahjoub Jabli*

Department of Textile Materials and Process Research (ENIM), National School of Engineers of Monastir, Tunisia

*Corresponding author: Mahjoub Jabli, Department of Textile Materials and Process Research (ENIM), National School of Engineers of Monastir, Monastir 5000, Tunisia

Submission: 📅 November 03, 2017; Published: 📅 January 09, 2018

Abstract

Natural dyes are considered as promising choice to develop green textile dyeing process. In this framework, different parts of various plants constitute potential sources of natural dyes that are characterized by their easy availability and abundant nature. In this present work, *Tradescantia pallida purpurea* and *Cynomorium coccineum* are considered, for the first time, as sources of natural dyes for textile dyeing. Their prepared aqueous methanolic extracts were chemically characterized for Their Total Phenolic Contents (TPC), Total Flavanoid Contents (TFC) and for their ability to quench reactive species through the DPPH* radical scavenging activity. The results gleaned from the dosage experiments of TPC and TFC values are found, to be 52mg GAE/g extract and 4.05mg QE/g extract for the methanolic *Cynomorium coccineum* extract. However, in the case of *Tradescantia pallida purpurea*, the TPC and TFC values are equal to 33mg GAE/g extract and 4.88mg QE/g extract. These results demonstrate that the polyphenolic compounds are the most important functional components found in the studied plants. These aqueous extracts are, also, rich with flavonoids which are generally known for their coloring power when they are applied to textile materials. *Tradescantia pallida purpurea* gives a pH dependent color. It is red at pH3 and yellow at pH8. However, the resulted dye from *Cynomorium coccineum* is blue.

Keywords: *Tradescantia pallida purpurea*; *Cynomorium coccineum*; TPC; TFC; IC50

Introduction

The consumption of the natural products has improved due to the global development of maintaining good health and reducing the risk of disease [1-3]. In this setting, many plants were studied. As example, *Tradescantia pallida purpurea* has been shown to have spatial and seasonal bio monitoring characteristics for metal emissions from vehicle pollution [4]. The antho cyanins unpurified crude extracted from this plant was, also, investigated with regards to their preliminary spectroscopic and thermo-optical characterization [5]. Several species of the genus *Tradescantia* belonging to the *Commelinaceae* family are, additionally, used in ethnobotany and used as medicinal and ornamental purposes [6,7].

Another plant called *Cynomorium coccineum* L. is a non-photosynthetic plant, spread over the south of Spain to the southern Italian coast, Sardinia, Sicily, Malta and from the West African coast to North African coast (The Canary Islands to Tunisia) [8-10]. It belongs to the family of *Balanophoraceae*. It is a blackish red leafless root parasitic plant [11]. It is known as a traditional medicine. It is valorized as a tonic and aphrodisiac [12] and it is reported to enclose a hypo tensive effect [13]. Hence, the majority of the studies have focused on the biological properties and it has performed with aqueous and ethanol extracts [14-17]. In this framework, the works of Antonella Rosa et al. [18] dealt with the study of the composition

and effect on intestinal Caco-2 cell viability and lipid profile of fixed oil obtained from dried stems of the plant and oil isolation has been performed by supercritical fractioned extraction with CO₂. However, to our knowledge, there is no data which was concentrated on the investigation of the extraction of dyes from *Tradescantia pallida purpurea* and *Cynomorium coccineum* fractions. Herein, for the first time, our study is carried out for the extraction of dyes from these plants. Their aqueous extract is characterized for their TFC, TPC values and the antioxidant activity.

Materials and Methods

Chemicals and materials

The fractions of *Tradescantia pallida purpurea* and *Cynomorium coccineum* (Figure 1) were collected from the region of Monastir Tunisia during 15 Mars-Avril. Chemicals such as Folin-Ciocalteu's reagent, gallic acid and 2, 2-diphenyl-1-picrylhydrazyl (DPPH) were purchased from Sigma-Aldrich. All other solvents and chemicals (Methanol, etc.) used in this study were of analytical grade [19].

Preparation of *Tradescantia pallida purpurea* and *Cynomorium coccineum* extracts

Fractions of *Tradescantia pallida purpurea* and *Cynomorium coccineum*, with weighted fresh mass, were extracted with absolute

methanol for a period of 72h at room temperature. After filtration, the fractions were evaporated using vacuum evaporator® apparatus (Figure 2a) and the extracted fraction were, diluted with distilled water and used for the dyeing experiments. *Tradescantia*

pallida purpurea gives a pH dependent color. It is red at pH₃ and yellow at pH 8. However, the resulted dye from *Cynomorium coccineum* is blue (Figure 2).



Figure 1: Photos of *Tradescantia pallida purpurea* and *Cynomorium coccineum* grown throughout the region of Monastir, Tunisia.

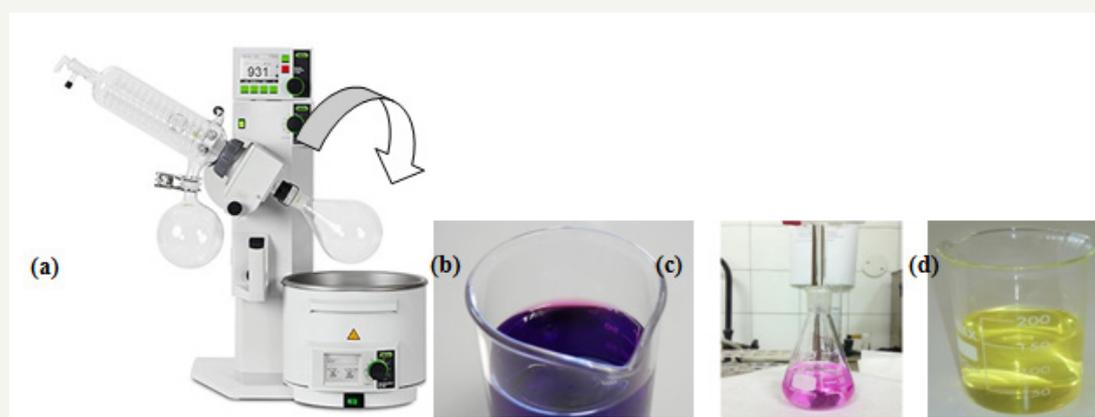


Figure 2: (a) Evaporator® apparatus, (b) dye extracted from *Cynomorium coccineum* fraction (c) dye extracted from *Tradescantia pallida purpurea* at pH=3 and (d) dye extracted from *Tradescantia pallida purpurea* at pH= 8.

The TFC value of *Tradescantia pallida purpurea* and *Cynomorium coccineum* methanolic extracts was determined based on the method of Sun et al. [20] with some modifications. Briefly, 20 μ L of the methanolic extract (1mg/m L) or quercetin (0.6-1mg/m L) was mixed with 30 μ L of NaNO₂ (5%). After 6min, 50 μ L of AlCl₃ (10%) was added and the resulting mixture was allowed to be kept for another 5min. To the above mixture, 100 μ L of NaOH (10%) was added and it has been incubated at room temperature for 15min. The absorbance was measured at 510nm and TFC value was expressed as Quercetin Equivalent (QE).

Free radical scavenging activity

The radical scavenging assay 1,1-diphenyl-2-picrylhydrazyl (DPPH) was carried out according to the method described by Yu et al. [21]. Briefly, a volume of 1mL of the as-prepared methanolic extract with different concentrations was mixed with 1mL of DPPH solution (0.1mM in ethanol). This mixture was incubated at room temperature for 30min, and the absorbance was measured at

517nm. The concentration required to scavenge 50% of DPPH* was determined based on the ascorbic acid calibration curve.

Results and Discussion

Dosage of TPC and TFC

Table 1: TPC, TFC, and IC50 determined for *Tradescantia pallida purpurea* and *Cynomorium coccineum* methanolic extracts.

Plants	TPC (Mg QE/G Extract)	TFC (Mg GAE/G Extract)	IC50
<i>Tradescantia pallida purpurea</i>	33	4.88	3.7
<i>Cynomorium coccineum</i>	52	4.05	7.5

The results of the dosage experiments of TPC and TFC values are 52mg GAE/g extract and 4.05mg QE/g extract for the *Methanolic cynomorium coccineum* extract (Table 1). However, in the case of *Tradescantia pallida purpurea*, the values are equal to 33mg GAE/g extract and 4.88mg QE/g extract. This table demonstrated that

the poly phenolic compounds are the most important functional components found in the studied extracts. These aqueous extracts are, also, rich with flavonoids which are generally known for their coloring power when they are applied to textile materials [22].

DPPH radical scavenging assays

The ability of *Tradescantia pallida purpurea* and *Cynomorium coccineum* fractions to quench reactive species by hydrogen donation was measured through the DPPH* radical scavenging

activity assay [21]. The evolution of DPPH scavenging activity (%) against the studied *Tradescantia pallida purpurea* and *Cynomorium coccineum* methanolic fractions extract concentration is depicted in Figure 3. IC50 values were deduced as 3.7mg/mL and 7.5mg/mL, respectively, for *Tradescantia pallida purpurea* and *Cynomorium coccineum*. These values are more important compared to Quercetin (0.064mg/mL). As, also, observed, the aqueous extract showed a concentration-dependent DPPH* radical scavenging activity.

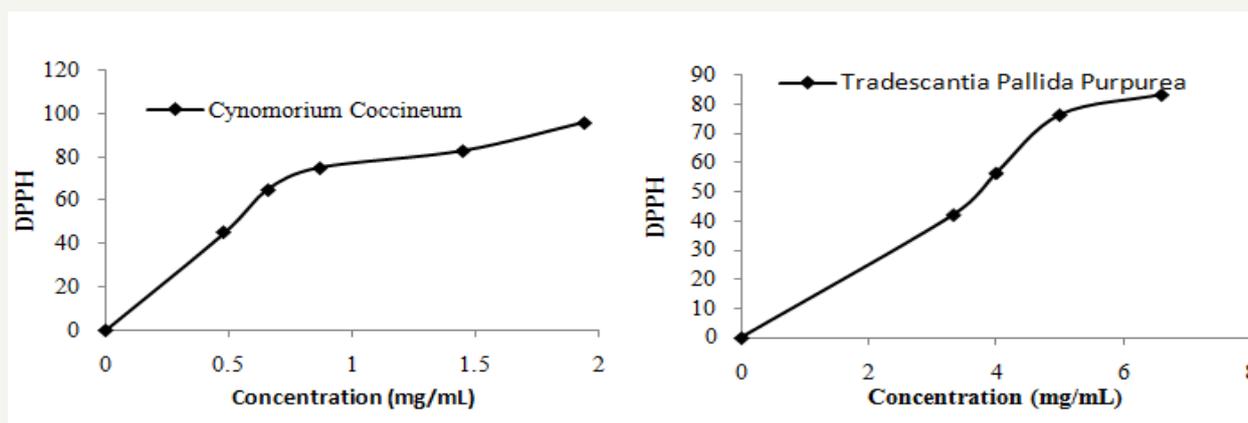


Figure 3: The ability of *Tradescantia pallida purpurea* and *Cynomorium coccineum* extracts to quench reactive species through the DPPH* radical scavenging activity assay.

Conclusion

To sum up, the methanolic extracts of *Tradescantia pallida purpurea* and *Cynomorium coccineum* were chemically characterized for their total phenolic contents, total flavanoid contents and for their ability to quench reactive species through the DPPH* radical scavenging. Data revealed that the dosage experiments of TPC values allows us to conclude that the polyphenolic compounds are the most important functional components found in the two studied plants. These aqueous extracts are, also, rich with flavonoids which are generally known for their coloring power for textile materials. Further works will be extended for the investigation of the full dyeing properties of the textile clothing using these studied methanolic extracts. The extraction process will also be optimized using various sorts of solvents.

References

1. Teresa SP, Sanchez BMT (2007) Anthocyanins: from plant to health. *Phytochemistry Reviews* 7(2): 281-299.
2. Suwalsky M, Vargas P, Avello M, Villena F, Sotomayor CP (2008) Human erythrocytes are affected in vitro by flavonoids of *Aristolochia chilensis* (Maqui) leaves. *Int J Pharm* 363: 85-90.
3. Kayesh E, Shangguan L, Korir NK, Sun X, Bilkish N, et al. (2013) Fruit skin color and the role of anthocyanin. *Acta Physiol Plant* 35(10): 2879-2890.
4. Santos APM, Segura Muñoz SI, Nadal M, Schuhmacher M, Domingo JL, et al. (2015) Traffic-related air pollution biomonitoring with *Tradescantia pallida* (rose) Hunt. cv. *purpurea* Boom in Brazil. *Environ Monit Assess* 187(2): 39.
5. Vanessa M, Sthanley R, Jaqueline OBT, Andrea A, Djalimir NM, et al. (2016) Preliminary spectroscopic and thermo-optical characterization of anthocyanin unpurified crude extracted from *Tradescantia pallida Purpurea*. *Dyes and Pigments* 135: 57-63.
6. Jing P, Bomser JA, Schwartz SJ, He J, Magnuson BA, et al. (2008) Structure-function relationships of anthocyanins from various anthocyaninrich extracts on the inhibition of colon cancer cell growth. *J Agric Food Chem* 56(20): 9391-9398.
7. Baublis AJ, Berber Jiménez MD (1995) Structural and conformational characterization of a stable anthocyanin from *Tradescantia pallida*. *J Agric Food Chem* 43(3): 640-646.
8. Abd el Rahman HA, el Badry AA, Mahmoud OM, Harraz FA (1999) The effect of the aqueous extract of *Cynomorium coccineum* on the epididymal sperm pattern of the rat. *Phytother* 13(3): 248-250.
9. Dharmananda S (2011) *Cynomorium*-Parasitic plant widely used in traditional medicine.
10. Duke JA, Duke PAK, du Cellier JL (2008) *Duke's Handbook of Medicinal Plants of the Bible*. (1st edn), CRC Press, Boca Raton, FL, USA, p. 552
11. Heestra H, Al Hassan H, Minwer F (1990) *Plants of the Northern Saudi Arabia, an Illustrated Guide*. Range and Animal Development Centre Publications, Skaka, Saudi Arabia, pp. 170-171.
12. Ageel AM, Mossa JS, Tariq M, Al Yahya MA, Al Said MS (1987) *Saudi Plants Used in Folk Medicine*. KACST, Riyadh, Saudi Arabia, p. 24.
13. Ikram M, Dar MS and Fakouhi T (1978) Hypotensive agent from *Cynomorium coccineum*. *Pahlavi Med J* 9(2): 167-181.
14. Abd El Rahman HA, El Badry AA, Mahmoud OM, Harraz FA (1999) The effect of aqueous extract of *C. coccineum* on the epididymal sperm pattern of the rat. *Phytother* 13(3): 248-250.
15. Al Qarawi AA, Abdel Rahman HA, El Badry AA, Harraz F, Razig NA, et al. (2000) The effect of extracts of *Cynomorium coccineum* and *Withania somnifera* on gonadotrophins and ovarian follicles of immature wistar rats. *Phytother* 14(4): 288-290.



16. Rached W, Benamar H, Bennaceur M, Marouf A (2010) Screening of the antioxidant potential of some Algerian indigenous plants. *J Biol Sci* 10(4): 316-324.
17. Abdel Magied EM, Abdel Rahman HA, Harraz FM (2001) The effect of aqueous extracts of *Cynomorium coccineum* and *Withania somnifera* on testicular development in immature Wistar Rats. *J Ethnopharmacol* 75(1): 1-4.
18. Antonella R, Antonio R, Alessandra P, Angela A, Paola S (2012) Chemical composition and effect on intestinal Caco-2 cell viability and lipid profile of fixed oil from *Cynomorium coccineum* L. *Food and Chemical Toxicology* 50(10): 3799-3807
19. Bursal E, Gülçin İ (2011) Polyphenol contents and *in vitro* antioxidant activities of lyophilized aqueous extract of kiwi fruit (*Actinidiadelicosa*). *Food Research International* 44: 1482-1489.
20. Sun L, Zhang J, Lu X, Zhang L, Zhang Y (2011) Evaluation to the antioxidant activity of total flavonoids extract from persimmon (*Diospyros kaki* L.) leaves. *Food Chem Toxicol* 49(10): 2689-2696.
21. Yu L, Zhao M, Wang JS, Cui C, Yang B, et al. (2008) Antioxidant, immunomodulatory, and anti-breast cancer activities of phenolic extract from pine (*Pinus massoniana* Lamb.) bark. *Innovative Food Science and Emerging Technologies* 9(1): 122-128.
22. Nouredine B, Wafa H, Manel BT, Maria TPA, Mohamed FM (2017) Sustainability issues of ultrasonic wool dyeing with grape pomace colourant. *Natural Product Research* 31(14): 1655-1662.



Creative Commons Attribution 4.0
International License

For possible submissions Click Here

[Submit Article](#)

**Your subsequent submission with Crimson Publishers
will attain the below benefits**

- High-level peer review and editorial services
- Freely accessible online immediately upon publication
- Authors retain the copyright to their work
- Licensing it under a Creative Commons license
- Visibility through different online platforms
- Global attainment for your research
- Article availability in different formats (**Pdf, E-pub, Full Text**)
- Endless customer service
- Reasonable Membership services
- Reprints availability upon request
- One step article tracking system