

Recent Advances in FRET Technology for Medical Applications

Ling Jiang, Ping Li, Ying Jiang, Zhitao Hou, Tianzhu Guo, Ping Liu, Yuting Ji, Yan Qi, XinYue Zhang, Yuan Tian, XiaoxiaoQi, Ruonan Zheng and Yun Bai*

School Basic Medical Science, Heilongjiang University of Chinese Medicine, Heilongjiang University of Chinese Medicine, P.R. China

Abstract

Förster resonance energy transfer (FRET) has emerged as a pivotal biophysical tool in medical research, enabling high-sensitivity detection of molecular interactions and dynamic processes. Recent technological advancements have significantly expanded FRET applications in cell biology and drug discovery, driving profound insights into disease mechanisms and therapeutic innovation. Despite its transformative potential, FRET implementation faces persistent challenges including signal intensity limitations, background noise interference, and experimental design constraints. This review synthesizes cutting-edge developments in FRET technology, critically evaluates its current medical applications, addresses key methodological challenges, and outlines future research trajectories to advance biomedical investigation.

Keywords: FRET; Biomedicine; Bioimaging; Cell biology; Drug discovery

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*Corresponding authors: Yun Bai, School Basic Medical Science, Heilongjiang University of Chinese Medicine, Heilongjiang University of Chinese Medicine, 24 Heping Road, Harbin, 150040, P.R. China

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Introduction

FRET technology leverages distance-dependent energy transfer between fluorescent molecules (donor and acceptor) to detect biomolecular interactions with nanometerscale spatial precision (<10nm). This technique has become indispensable for monitoring real-time cellular signaling, elucidating disease pathways, and facilitating high-throughput drug screening. Recent innovations have broadened FRET's utility across diverse contexts from mapping single-molecule dynamics to profiling network-level interactions in complex biological systems [1]. Notable progress includes FRET-based monitoring of intracellular signaling cascades that reveal adaptive cellular responses to stimuli [2], identification of pathogenic protein interactions in cancer and neurodegenerative disorders [3], and accelerated drug candidate evaluation through multiplexed screening platforms [4]. This review examines fundamental principles and emerging applications of FRET technology across medical domains, highlighting its transformative potential in addressing critical biomedical challenges.

Fundamental principles and biomedical significance of FRET

- Mechanism and Requirements FRET efficiency (E) follows Förster's equation E = $1/[1+(R/R_0)^6]$, where R represents donor-acceptor distance and R_0 denotes the characteristic distance at 50% energy transfer. Optimal FRET occurs when spectral overlap exceeds 30%, dipole orientations are favorable, and interfluorophore distance remains within 1-10nm. These characteristics position FRET as a powerful "molecular ruler" for quantifying biomolecular interactions1.
- B. Advantages in Biomedical Research FRET provides unparalleled capabilities for livecell dynamic monitoring with millisecond temporal resolution and single-molecule

sensitivity. Its compatibility with fluorescence lifetime imaging microscopy (FLIM) enables background-free quantification [5,6]. Recent advances including time-resolved detection and multiphoton excitation have further enhanced signal-to-noise ratios (>500:1) and tissue penetration depth (up to $800\mu m$) [7].

FRET in cell biology

Dynamic monitoring of cellular signaling: Integration of FRET with two-photon fluorescence lifetime imaging microscopy (2P-FLIM) enables spatiotemporal mapping of kinase activation and second-messenger dynamics in thick tissues [7]. For instance, Rac1 GTPase activity gradients in migrating cells have been quantified using EGFP-mCherry FRET biosensors with subcellular resolution (±0.1 FRET efficiency units) [8-20].

Protein interaction analysis: Dual-labeling strategies with fluorescent proteins (e.g., CFP-YFP) reveal protein complex stoichiometry and dissociation constants *in vivo*. This approach has identified allosteric changes in $\beta 2$ -adrenergic receptors within 500ms of agonist stimulation.

Molecular diagnostics innovation: smFRET, a Single-molecule resolution achieves 10⁻¹⁸M detection limits for nucleic acid dynamics and CRISPR-Cas activity monitoring [21,22]. TR-FRET, Europium chelate donors with time-gated detection reduce autofluorescence interference by 30-fold for cytokine storm diagnostics [23]. Quantum Dot (QD)-FRET, Water-soluble CdTe/ZnS core-shell QDs functionalized with 3-mercaptopropionic acid exhibit quantum yields >80% for quantitative metabolite sensing [24].

FRET in drug discovery

- A. Target Validation TR-FRET binding assays screen interactions at physiological concentrations, outperforming ELISA in precision and throughput [25].
- B. Screening and Mechanism Profiling Advanced FRET biosensors enable real-time drug activity assessment: Genetically encoded mNeonGreen-HaloTag systems track receptor dimerization kinetics Carbon nanotube-QD hybrids detect serum PD-L1 at 0.1pM in 5 minutes Lanthanide nanoparticle probes guide neurosurgical interventions with <50μm precision [26,27]. Integrated genotyping (e.g., LightSNiP + TR-FRET) identifies DPYD 6 variants predictive of fluoropyrimidine toxicity [28].</p>

Conclusion

FRET is undergoing rapid development in medical applications, extending its influence to critical domains including cell biology and drug discovery. Through non-invasive means, FRET enables real-time monitoring of molecular interactions and dynamic processes within cells, significantly advancing our understanding of biological mechanisms. However, as FRET gains widespread adoption, researchers face challenges in balancing divergent perspectives. On the one hand, studies demonstrate FRET's superiority in monitoring molecular interactions and dynamic changes, providing high-resolution, real-time intracellular information. On the other hand,

concerns exist regarding its accuracy in complex biological systems, where signal interference and background noise may compromise result reliability. To reconcile these views, future research must prioritize standardization and optimization of FRET technology for precise application across diverse biological environments.

Continual advancements in FRET bring both challenges and opportunities. Innovations in probe development, imaging technology, and data analysis methods are expanding FRET's potential. These improvements will enhance its value in basic research and accelerate clinical translation for early disease diagnosis and personalized therapies.

Future studies should persistently explore novel FRET applications and integrate it with complementary bioimaging technologies to further medical research. Through interdisciplinary collaboration, researchers can achieve comprehensive understanding of complex biological systems, providing robust support for clinical practice. In summary, while FRET faces significant challenges, its promising future in medicine holds undeniable potential to profoundly impact biomedical research.

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Conflicts of interest

There are no conflicts to declare.

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References

- Fang C, Huang Y, Zhao Y (2023) Review of FRET biosensing and its application in biomolecular detection. Am J Transl Res 15 (2): 694-709.
- Nakashima S, Toyama A, Sugiyama H, Aoki K, Goto Y (2025) Capturing CDKs in action: Live-cell biosensors pioneer the new frontiers in cell cycle research. Cell Struct Funct 50(1): 77-90.
- Yu H, Feng R, Chen F, Wu Z, Li D, et al. (2024) Rapid FRET assay for the early detection of alpha-synuclein aggregation in parkinson's disease. ACS Chem Neurosci 15(7): 1378-1387.
- Kelly T, Yang X (2024) Application of fluorescence- and bioluminescencebased biosensors in cancer drug discovery. Biosensors (Basel) 14 (12): 570.
- Zlotnikov ID, Savchenko IV, Kudryashova EV (2023) Fluorescent probes with forster resonance energy transfer function for monitoring the gelation and formation of nanoparticles based on chitosan copolymers. J Funct Biomater 14(8): 401.
- Vu CQ, Arai S (2023) Quantitative imaging of genetically encoded fluorescence lifetime biosensors. Biosensors (Basel) 13(10): 939.

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- Long Y, Stahl Y, Weidtkamp-Peters S, Postma M, Zhou W, et al. (2017) In vivo FRET-FLIM reveals cell-type-specific protein interactions in Arabidopsis roots. Nature 548(7665): 97-102.
- Liao JM, Wang YT, Chen CL (2011) Computer simulation to investigate the FRET application in DNA hybridization systems. Phys Chem Chem Phys 13(21): 10364-10371.
- Song Y, Madahar V, Liao J (2011) Development of FRET assay into quantitative and high-throughput screening technology platforms for protein-protein interactions. Ann Biomed Eng 39(4): 1224-1234.
- 10. Song Y, Liao J (2012) Systematic determinations of SUMOylation activation intermediates and dynamics by a sensitive and quantitative FRET assay. Mol Biosyst 8(6): 1723-1729.
- Song Y, Rodgers VG, Schultz JS, Liao J (2012) Protein interaction affinity determination by quantitative FRET technology. Biotechnol Bioeng 109(11): 2875-2883.
- Liu Y, Liao J (2013) Quantitative FRET (Forster Resonance Energy Transfer) analysis for SENP1 protease kinetics determination. J Vis Exp (72): e4430.
- 13. Jiang L, Saavedra AN, Way G, Alanis J, Kung R, et al. (2014) Specific substrate recognition and thioester intermediate determinations in ubiquitin and SUMO conjugation cascades revealed by a high-sensitive FRET assay. Mol Biosyst 10(4): 778-86.
- 14. Malik-Chaudhry HK, Saavedra A, Liao J (2014) A linker strategy for trans-FRET assay to determine activation intermediate of NEDDylation cascade. Biotechnol Bioeng 111(7): 1288-1295.
- 15. Liao JY, Song Y, Liu Y (2015) A new trend to determine biochemical parameters by quantitative FRET assays. Acta Pharmacol Sin 36(12): 1408-1415.
- 16. Wiryawan H, Dan K, Etuale M, Shen Y, Liao J (2015) Determination of SUMO1 and ATP affinity for the SUMO E1by quantitative FRET technology. Biotechnol Bioeng 112(4): 652-658.
- 17. Jiang L, Xiong Z, Song Y, Lu Y, Chen Y, et al. (2019) Protein-protein affinity determination by quantitative FRET quenching. Sci Rep 9(1): 2050.

- 18. Way G, Xiong Z, Wang G, Dai H, Zheng S, et al. (2020) A novel SUMOylation site in the influenza a virus NS1 protein identified with a highly sensitive FRET assay. J Biotechnol 323: 121-127.
- Liao J, Madahar V, Dang R, Jiang L (2021) Quantitative FRET (qFRET) technology for the determination of protein-protein interaction affinity in solution. Molecules 26(21): 6339.
- 20. Liu Y, Zhang F, Jiang L, Perry JJP, Zhao Z, et al. (2021) Product inhibition kinetics determinations - Substrate interaction affinity and enzymatic kinetics using one quantitative FRET assay. Int J Biol Macromol 193(Pt B): 1481-1487.
- Yang Z, Xu H, Wang J, Chen W, Zhao M (2021) Single-molecule fluorescence techniques for membrane protein dynamics analysis. Appl Spectrosc 75(5): 491-505.
- 22. Groves D, Hepp C, Kapanidis AN, Robb NC (2023) Single-molecule FRET for virology: 20 years of insight into protein structure and dynamics. Q Rev Biophys 56: e3.
- 23. Morrison LE (1988) Time-resolved detection of energy transfer: Theory and application to immunoassays. Anal Biochem 174(1): 101-120.
- 24. Agbulut MSB, Elibol E, Cadirci M, Demirci T (2025) Fluorescent CdTe/ ZnS core/shell quantum dots for sensitive metabolite detection in real samples. J Fluoresc.
- 25. Blay V, Tolani B, Ho SP, Arkin MR (2020) High-Throughput Screening: today's biochemical and cell-based approaches. Drug Discov Today 25(10): 1807-1821.
- 26. Tarpley M, Caligan TB, Onyenwoke RU, Williams KP (2021) Optimization and validation of a DYRK1A TR-FRET assay for high-throughput screening. MethodsX 8: 101383.
- 27. Wu Q, Lin W, Li ZM, Rankovic Z, White SW, et al. (2021) A protocol for high-throughput screening of histone lysine demethylase 4 inhibitors using TR-FRET assay. STAR Protoc 2(3): 100702.
- 28. Montrasio C, Cheli S, Clementi E (2023) Pharmacogenetic practice of anticancer drugs: Multiple approaches for an accurate and comprehensive genotyping. Pharmgenomics Pers Med 16: 739-746.

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