Influence of Wall Material Composition on Microencapsulation Efficiency of Cold Pressed Pumpkin Seed Oil by Freeze-Drying

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Abstract

The main goal of this research was to investigate the effects of wall materials on microencapsulation efficiency of pumpkin seed oil by freeze-drying. Different ratios of maltodextrin (MD), xanthan gum (XG), gum arabic (GA), whey protein concentrate (WPC 80), casein and, lecithin combinations were used as wall materials to encapsulate cold pressed pumpkin seed oil. Microencapsulation efficiency values of microcapsules ranged between 12.36-34.51%. The maximum microencapsulation efficiency was achieved using maltodextrin, gum arabic, whey protein concentrate, and lecithin combination.

Introduction

Encapsulation is a food preservation method where sensitive compounds, known as “core material”, can be entrapped within a matrix that is called “wall material” [1]. Capsules of which sizes range between 3-800µm are called microcapsules, while particle sizes of Nano capsules range between 10-1000nm [2]. The most widely used microencapsulation techniques are spray drying, spray cooling, extrusion, fluidized bed coating, freeze-drying (lyophilization), coacervation, cocrystallization, interfacial or in-situ polymerization [3,4]. Although its high production costs, freeze-drying is a unique technology for heat sensitive core materials. The well-known reasons of encapsulation are protection of core material against environmental factors such as oxygen, light or heat and hence, extending the shelf life of product; improvement of core materials’ handling and storage properties; controlled release of core materials’ handling and storage properties; controlled release of core materials; and/or masking of off-odour and off-taste [1,5].

Main compounds that need to be encapsulated are flavours, antioxidants, antimicrobials, bioactive peptides, minerals, vitamins, bioactive lipids, probiotics and dietary fibres [6]. Various carbohydrates (starch, cellulose, gum Arabic, carrageenan, alginate, xanthan, chitosan etc.), proteins (whey protein, casein, gluten, gelatin etc.) and lipids (phospholipids, fatty acids, waxes etc.) may be used as wall materials in encapsulation processes [7]. The main advantages of maltodextrins are serving as an efficient barrier against oxidation, neutral taste, low cost, high solubility in water and formation of low viscosity solutions even at high concentrations. However, due to lack of emulsifying capacity of maltodextrins, they should be used in combination with surface active wall materials such as gum arabic [8,9]. In encapsulation processes, milk proteins, whey and casein, improve stability of food emulsions by their high film-forming and emulsifying properties [10]. It should be kept in mind that whey acts as a more effective emulsifier in oil-in-water emulsions than casein and sodium caseinate [11]. Surface active activity of gum arabic is attributed to its low protein content [12]. Nevertheless, its high price and limited availability require its incorporation into other wall materials [13]. Lecithin may be added to wall material combinations to enhance wettability and dispersibility properties of microcapsules containing oil or fat [14].

Pumpkin (Cucurbita pepo subsp. pepo var. Styriaca) seed oil is a specialty oil that contains substantial amounts of unsaturated fatty acids, mainly linoleic and oleic acids, squalene, phytosterols, tocopherols and tocotrienols [15-17]. All of these are accepted as bioactive compounds due to their health promoting properties. However, pumpkin seed oil rich in unsaturated fatty acids is susceptible to oxidation. Microencapsulation of pumpkin seed oil by freeze-drying may protect the oil against environmental conditions and improve the oxidative stability of oil. The objective of this study is to investigate the effects of wall materials on encapsulation efficiency of cold pressed pumpkin seed oil by freeze-drying.

Materials and Methods

Material

Pumpkin seeds were purchased from Yüredi Gıda San. Tic. Ltd. Şti. (Konya, Turkey). Then, oil of the seeds was obtained by using a Cold-pressing machine (Toper Machine, Izmir, Turkey). The temperature of the machine maintained at 40 °C during operation. Next, oils were centrifuged (Hettich Universal 32R, Germany) at 4000rpm for 20 min at 15 °C. Finally, oils were transferred...
to amber coloured bottles under nitrogen and stored at 4 °C until further use. Cold pressed pumpkin seed oil with the following physicochemical properties was used as core material: Lovibond R: 39; Lovibond Y: 19; refractive index at 20°C: 1.45545; free fatty acids: 0.31% oleic acid; iodine value: 101.31g/100g; peroxide value: 4.03meq O2/kg; total chlorophyll content: 2.86mg phophetytin a/kg; total carotenoid content: 5.42mg lutein/kg, DPPH-scavenging activity: 5.50mM Trolox equivalent/100g; total phenolic content: 13.25mg gallic acid equivalent/100g; 37.03% oleic acid, 43.67% linoleic acid, 11.11% palmitic acid, 6.70% stearic acid; 180.89mg γ-tocopherol/100g and 91.30mg α-tocopherol/100g.

**Table 1:** Compositions of emulsions.

<table>
<thead>
<tr>
<th>Run</th>
<th>MD (g)</th>
<th>XG (g)</th>
<th>GA (g)</th>
<th>WPC 80 (g)</th>
<th>Casein (g)</th>
<th>Lecithin (g)</th>
<th>Pumpkin Seed Oil (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Run-1</td>
<td>0.8</td>
<td>-</td>
<td>0.2</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>1</td>
</tr>
<tr>
<td>Run-2</td>
<td>0.8</td>
<td>0.5</td>
<td>-</td>
<td>-</td>
<td>0.2</td>
<td>-</td>
<td>1</td>
</tr>
<tr>
<td>Run-3</td>
<td>0.8</td>
<td>-</td>
<td>0.5</td>
<td>0.2</td>
<td>-</td>
<td>-</td>
<td>1</td>
</tr>
<tr>
<td>Run-4</td>
<td>0.8</td>
<td>-</td>
<td>0.5</td>
<td>-</td>
<td>0.2</td>
<td>-</td>
<td>1</td>
</tr>
<tr>
<td>Run-5</td>
<td>0.8</td>
<td>-</td>
<td>0.5</td>
<td>0.2</td>
<td>-</td>
<td>0.002</td>
<td>1</td>
</tr>
<tr>
<td>Run-6</td>
<td>1.6</td>
<td>-</td>
<td>0.5</td>
<td>0.2</td>
<td>-</td>
<td>0.002</td>
<td>1</td>
</tr>
<tr>
<td>Run-7</td>
<td>0.8</td>
<td>-</td>
<td>0.5</td>
<td>0.8</td>
<td>-</td>
<td>0.002</td>
<td>1</td>
</tr>
</tbody>
</table>

**Emulsion preparation**

The wall materials (MD, XG, GA, WPC 80, casein and, lecithin) were selected according the previous similar studies. The composition of each run was summarized in Table 1. Wall materials were separately dissolved in distilled water by mixing with magnetic stirrer (Wisd Wise Stir MSH-20A, Germany) at 1500rpm for 10 min. Then, each wall material solution was combined in a stainless-steel beaker and pumpkin seed oil was added to this mixture. Emulsions were formed using a rotor stator homogenizer (Velp OV-5, Italy) at 22,000rpm for 7min. Obtained emulsions were frozen in petri dishes at -25 °C for 24h.

**Microencapsulation by freeze-drying**

Freeze-drying process was performed in a laboratory scale freeze dryer (Christ Alpha 2-4 LD, Germany) operating at -80 °C for 48h. After freeze-drying, pellets were ground into powder using a knife mill (Retsch Grindomix GM200, Germany).

**Encapsulation efficiency**

Total and surface oil fractions of microcapsules were extracted according to the methods suggested by Velasco et al. [18]. Encapsulation efficiency was calculated using the following equation:

\[
\text{Encapsulation efficiency} (%) = \left(\frac{\text{Total oil} - \text{Surface oil}}{\text{Total oil}}\right) \times 100
\]

**Statistical analysis**

Experiments were performed in duplicate. Data were subjected to one-way analysis of variance (ANOVA) in SAS Ver 8.2 and compared by Fisher's LSD test method at p<0.05. Results were presented as mean ± standard deviation.

**Results and Discussion**

**Table 2:** Free oil contents and microencapsulation efficiency values of runs.

<table>
<thead>
<tr>
<th>Run</th>
<th>Surface Oil (%)</th>
<th>Microencapsulation Efficiency (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Run-1</td>
<td>24.20±0.91*</td>
<td>16.32±1.64*</td>
</tr>
<tr>
<td>Run-2</td>
<td>24.29±0.60*</td>
<td>12.36±0.69*</td>
</tr>
<tr>
<td>Run-3</td>
<td>16.35±0.63*</td>
<td>33.62±0.72*</td>
</tr>
<tr>
<td>Run-4</td>
<td>19.52±0.98*</td>
<td>28.67±0.72*b</td>
</tr>
<tr>
<td>Run-5</td>
<td>15.59±0.30*</td>
<td>34.51±0.51*</td>
</tr>
</tbody>
</table>

Run-6 21.21±0.41*   19.54±0.55*  
Run-7 20.22±0.71*   22.04±0.51*

Microencapsulation efficiency is the best indicator of the success of any encapsulation process. As the amount of oil found on the surface of microspule decreases, the amount of encapsulated oil and correspondingly, encapsulation efficiency increases. Surface oil contents of the microcapsules and microencapsulation efficiency of each run were given in Table 2. Microencapsulation efficiency of runs ranged between 12.36-34.51%. Higher microencapsulation efficiency values by freeze-drying were reported for fish oil (29.40-81.60%) [19]; flaxseed oil (32.68-59.63%) [20]; olive oil (36.90-69.09%) [21], and rosemary essential oil (69.90-96.14%) [22] previously. Although all of these oils were encapsulated using freeze-drying, extremely different microencapsulation efficiency values reported by various researchers may be explained by various type and ratio of wall materials used; ratio of core-to-wall material and type of homogenizer (high shear rotor-stator homogenizer, ultrasonic homogenizer, microfluidizer, high pressure homogenizer). Effects of wall material composition on free oil amounts and microencapsulation efficiency of microcapsules were statistically significant (p<0.05). Due to its high viscosity, xanthan gum decreased the efficiency of mixing and homogenization of emulsions. Consequently, microcapsules containing xanthan gum (Run-1 and Run-2) had lower microencapsulation efficiency values than the corresponding microcapsules with gum arabic (Run-3 and Run-4). Moreover, surface active characteristic of gum arabic may contribute to higher stability of pumpkin seed oil-in-water emulsions. A stable emulsion inhibits oil droplet aggregation during freezing and provides increased microencapsulation efficiency. Addition of lecithin into
emulsion consisting of maltodextrin, gum arabic and whey protein concentrate (Run-3) caused a slight and statistically insignificant (p>0.05) increase in microencapsulation efficiency. Substitution casein for whey protein concentrate decreased microencapsulation efficiency of freeze dried microcapsules. However, our results contrast with the findings of Fäldt & Bergenståhl [23], who reported better encapsulation ability of sodium caseinate than whey protein for soybean oil microencapsulation [23]. Shu & Rosenberg [24] suggested that interaction between carbohydrate (maltodextrin) and protein (whey) may alter the emulsification properties of emulsions [24]. The changes observed when maltodextrin (Run-6) or whey protein concentrate (Run-7) amounts increased in the emulsion may be attributed to strength of protein-carbohydrate interactions explained by Shu & Rosenberg [24]. Moreover, differences between surface oil contents and microencapsulation efficiency of our protein-stabilized emulsions may be affected by instabilities like droplet coalescence occurred during freezing of emulsions [25]. Results obtained through our study verified the use of wall material combinations containing different ratios of maltodextrin, xanthan gum, gum arabic, casein, whey protein concentrate and lecithin for encapsulation of cold pressed pumpkin seed oil. Among all wall material combinations produced in this study, blend of maltodextrin + gum Arabic + whey protein concentrate + lecithin (Run-5) yielded the maximum encapsulation efficiency (34.51%).

Conclusion

This study demonstrated the applicability of different combinations of selected wall materials for cold pressed pumpkin seed oil microencapsulation by freeze-drying. Lower encapsulation efficiency values may be improved by optimizing concentration of each component serving as wall material and optimizing of core-to-wall material ratio or determination of suitable emulsion freezing conditions. Further studies will focus on improvement of microencapsulation efficiency of microcapsules and investigation of these microcapsules in terms of oxidative parameters such as peroxide value, p-alinidine value and TOTOX value. Moreover, another research could also be designed to investigate the effects of encapsulation process parameters on biologically active compounds of oils.

References