Background: Breast cancer is the most common among female, constituting about 18% of all female cancers, with 1.7 million new cases reported in the world each year. Recently some studies reported that approximately 18% of cancer cases can be linked to infectious agents including viruses particularly Human Papilloma Virus (HPV) and Epstein Barr virus (EBV).

Objective: Molecular detection of EBV and HPV types 16 and 18 in breast cancer patients in Khartoum State, Sudan.

Methods: Paraffin embedded blocks of tumor specimen from 70 Sudanese patients with breast cancer were collected from Omdurman teaching Hospital, Sudan, during the period from March to June 2018. PCR was used to investigate the presence of EBV and HPV type 16, 18 viruses in these specimens.

Results: The results show that eight out of 70 patients were positive for EBV virus (11.4%) Of these positive patients, 3(4.2%) were 30-50, 3(4.2%) were 51-80 and 2(2.8%) were 81-100 years old, respectively. Seven out of the 70 patients were positive for HPV type 18(10%). Of these positive patients, 5 (7.1%) were 30-50 years old and 2 (2.9%) were 50-80 years old. None of the patients were found positive for HPV type 16. No significant differences were found between age groups as regards infection by EBV or HPV type 18 viruses.

Conclusion: The incidence of EBV and HPV types 18 in breast cancer patients in Khartoum State was documented through the molecular detection of these two virus's DNA. Detection of EBV and HPV type18 using PCR was established. Generally, these findings are useful for future studies since there is little information available about of EBV and HPV types 16, 18 in Sudan.

Keywords: EBV and HPV16,18; Breast Cancer; Khartoum State; Sudan

Introduction

Breast cancer is a type of cancer originating from breast tissue, most commonly from the inner lining of milk ducts or the lobules that supply the ducts with milk [1]. The size, stage, rate of growth, and other characteristics of the tumor determine the kinds of treatment. Treatment may include surgery, drugs (hormonal therapy and chemotherapy), radiation and/or immunotherapy [2]. Many factors including radiation, chemicals and viruses, have been found to induce human cancer [3]. Viral factors are the most important class of the infectious agents associated with human cancers [4]. It was estimated that 17-20 % of the worldwide incidence of cancers was attributable to a viral etiology [5]. Infections with oncogenic viruses have been investigated as possible risk factors for a breast cancer aetiology including mouse mammary tumor virus (MMTV), Epstein-Barr virus (EBV) and human papilloma virus (HPV) especially types 16, 18, and 33; however, their prescise role is not clear. EBV was the first human virus to be directly implicated in carcinogenesis. Epstein Barr virus; a common infection affecting over 90% of the world’s population is one of the viruses that have some unclear and controversial points over its ability to trigger the development of certain tumors [6] such as Burkitt’s lymphoma, nasopharyngeal carcinoma, Hodgkin’s disease, gastric carcinoma and post-transplant lymphoproliferative disease [7].
The small untranslated RNAs EBER-1 and -2 are accumulated at high levels during all forms of latency and regulate apoptosis through different mechanisms which play a critical role in efficiency of EBV-induced growth transformation of primary B cells. EBER-1 interacts with the interferon-inducible protein kinase R (PKR), and inhibits its activation by double stranded RNAs, protecting infected cells from IFN-induced apoptosis [8]. However, Wu et al. [9] also reported that EBV-encoded RNA 2 (EBER2) but not EBER1 playing a critical role in EBV-induced B-Cell growth transformation. In Sudan, breast cancer is characterized by a geographically focal nature, early onset and aggressive course of the disease [10]. BRCA1, BRCA2, and p53 mutations are infrequent in Sudanese breast cancer patients. Epigenetic changes are suggested as alternative mechanisms to account for the minor contribution in the genetic alterations in three tumor suppressor genes, BRCA1, BRCA2, and p53, in both sporadic and familial breast cancer cases in Sudan [11].

Materials and Methods

Patient criteria and specimen collection

Paraffin embedded blocks of tumor specimens from 70 Sudanese patients with breast cancer were collected from Omdurman Teaching Hospital Sudan during the period from March to June 2018. The collected specimens were stored at room temperature until the test was performed.

Specimen deparaffinisation: This was carried out as recommended in the protocol using the DNA extraction kit (Acrogene, USA). The specimens were deparaffinized according to the protocol of the manufacturer DNA extraction procedure was carried out according to manufacturer’s protocol (Acrogene, USA).

DNA extraction

The procedure was carried out according to manufacturer’s protocol (Acrogene, USA).

Polymerase Chain Reaction (PCR) for detection of EBV: The PCR was performed using primers that are specific for the EBV (gB) conserved regions. The primers used consisted of forward primer SL1, 5-GGACCTCAAAGAAGGGGG-3 and the reverse primer SL3, 5-GCTCCTGGTCTTCCGCCTCC-3. The reaction was performed in a thermo cycling condition (10pg), 1L master mix, 1L extracted DNA and 12L distilled water. DNA was amplified as follow: 35 cycles of denaturation at 95°C for 10min, followed by 35 cycles of denaturation at 95°C for 45 sec, annealing at 60°C for 45sec and extension at 72°C for 60sec, with final extension at 72°C for 10min. The expected size of UTR gene amplicon was 80bp [12].

Polymerase Chain Reaction (PCR) for detection of HPV types 16 and 18: Samples were amplified separately for HPV 16 and 18 with a thermocycler (Techno, Japan), and primers specific for each viral type derived from previously published sequences (12,40). The sequence of primers for HPV 16 DNA were F, 5’TTTGGTTTACA CAT’TACAAG3’ (residues 7864 to 7885), and R, 5’TGTGC TGCTTTJ7’ATACTAAACCG3’ (residues 57 to 78), which generated a 119-bp product from the URR region. The sequence of primers for HPV 18 DNA were, F 5’GACACCTTAATGAAAAACGACGA3’ (residues 460 to 482), and R,5’CGTCGTTFGAGCTGTTCCTG3’ (residues 543 to 562), which amplified a 103-bp fragment from open reading frame E6. The reaction was performed in 25μL volume of PCR PreMix master mix (Intron Biotechnology, Korea). The volume included 5μL master mix, 1μL forward primer (10pg), 1μL reverse primer (10pg), 5μL extracted DNA and 12μL distilled water. DNA was amplified as follow: 35 cycles of denaturation at 94°C for 45sec, annealing at 54°C for HPV 16 or 58°C for HPV 18 for 1min 45sec and extension at 74°C for 30sec [13].

Agarose gel electrophoresis

10μL of amplified product was analyzed by gel electrophoresis in 2% agarose stained with 0.15% ethidium bromide and visualized by using UV gel documentation system (INGeNiuse Germany).

Statistical analysis

Collected data were analyzed using statistical package for social science (SPSS version 12.0). A p value of ≤ 0.05 was considered significant.

Result

Detection of EBV using PCR in breast cancer patients

Eight out of 70 patients were positive for EBV virus (11.4%). Of these positive patients, 3(4.2%) were 30-50, 3(4.2%) were 51-80 and 2(2.8%) were 81-100 years old respectively. Seven out of the 70 patients were positive for HPV type 18(10%). Of these positive patients, 3(4.2%) were (30-50), 3(4.2%) were (51-80) and 2(2.8%) were (81-100) years old, respectively as shown in (Table 1) but with no significant differences.

Detection of HPV types 16, 18 using PCR in breast cancer patients

Table 1: Detection of EBV, HPV types 16, 18 in breast cancer patients.

<table>
<thead>
<tr>
<th>Age Groups in Year</th>
<th>EBV Positive Results (%)</th>
<th>HPV Type 16 Positive (%)</th>
<th>HPV Type 18 Positive (%)</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>30-50</td>
<td>3 (4.5)</td>
<td>0 (0)</td>
<td>5 (7.6)</td>
<td>8(5.3)</td>
</tr>
<tr>
<td>50-80</td>
<td>3 (4.5)</td>
<td>0 (0)</td>
<td>2 (3.0)</td>
<td>5(3.3)</td>
</tr>
<tr>
<td>81-100</td>
<td>2 (3.0)</td>
<td>0 (0)</td>
<td>0 (0)</td>
<td>2(1.3)</td>
</tr>
<tr>
<td>Total</td>
<td>8 (12.1)</td>
<td>0 (0)</td>
<td>7 (10.6)</td>
<td>15(100)</td>
</tr>
</tbody>
</table>

* means significant difference

Note: No of patients in age group 30-50 is 43, in age group 50-80 is 20 and in 81-100 is 3.
Seven out of 70 patients were positive for HPV type 18(10%). Of these positive patients, 5 (7.1%) were 30-50 years old and 2 (2.9%) were 50-80 years old as shown in (Table 1). All the samples were negative for HPV type 16.

Discussion

Breast cancer is the most common malignancy among females and comprises about 18% of all cancers affecting them. About 1.7 million new cases are reported in the world each year. Based on the most global recent data, approximately 12.3% of women are diagnosed with breast cancer at a point of time during their life. Little is known about the viral causes of breast cancer and their epidemiology in Africa. Even much less is known about the epidemiology of viruses associated with breast cancer in Sudan in particular and in Africa in general. Our current study in Sudan is one of the few studies to report directly measured rates of EBV and HPV types 16, 18 associated with breast cancer patients in Sudan. Some studies have reported on the findings that EBV, HPV types 16, 18 play a role in the development of breast cancer including studies from Sudan [14-18], where this were reported in Sudan [16,19]. On the other hand, several studies failed to detect HPV positivity in breast carcinoma [20] even by studying HPV subtypes 6b, 11, 13, 16, 18, 30, 31, 32, 33, 45, and 51 in 95 women with breast cancer without detecting any of the subtypes [21] researched the HPV subtypes 6, 11, 16, and 18 in 13 IDC, 15 papillomas, and 15 papillary carcinomas cases, and there was no evidence of HPV infection [22].

When studying the prevalence of subtypes of high (16, 18, 33, and 45) and low risks (6, 11) in 50 breast cancers in women in France by PCR with the general primer GP5+/GP6+, where none of these subtypes were detected in both cases [23]. This may be due to the lack of standardized technique to detect the presence of HPV, since there are different types of primers for different HPV subtypes. False negatives and false positives may occur when PCR overestimates the association between HPV and breast cancer because it cannot indicate which types of cells the virus has infected. Contamination while handling the sample may be partially responsible for the high frequencies of HPV positivity that were reported in several papers. Other risk factors might affect the outcome of the results. For example, studies have shown that women under the age of 25 have a higher prevalence of HPV without detecting any of the subtypes [21] and without detecting any of the subtypes [22].

Storage of specimens may affect the results, since some researchers found that positive specimens became negative after being frozen for 3 months [25]. Demographic features and genetic backgrounds may also contribute to the geographic difference and HPV infection in breast cancer. The present study focused on the molecular detection of EBV and high-risk HPV types 16 and 18 in breast cancer patients in the Khartoum State. Elnoubi [19] was the first to detect HPV type 16,18 in breast cancer patients in Sudan. The author detected HPV genotype 16, in 21(31%), and HPV genotype 18, in 10(15%) of the tested patients. On the other hand, Adam detected EBV genome in 49 (53.3%) and 10 (11%) patients by LMP-1 and EBNA-4 PCR, respectively. Based on the results of the present study, findings of Elnoubi [19] and Adam AA [16] that suggest that, EBV, HPV type 18 infection may be common in Sudan. The present study in addition to the above-mentioned studies could serve as a baseline for future plans aiming at introducing the vaccine against EBV and HPV in the Sudan. Finally, these findings should highlight the need for the establishment in Sudan of rapid, sensitive, and specific diagnostic techniques (such as ones used here) to better understand the role played by various viruses in the etiology of breast cancer development in Sudan.

References


