

The Unsung Hero: The Role of *Agrobacterium* in Biodiversity

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Abstract

Broadly speaking, biodiversity refers to the variety of life on Earth that arises from the intricate and interdependent network of species, ecosystems, and genetic variation. Genetic recombination and mutation give birth to genetic variation even within species. Other factors that aid in environmental adaptation include genetic polymorphism, isolation of gene pools, local selection pressures, and environmental complexity. For example, many crop variations exist because of genetic variety, which enables the types to adapt to changing environmental conditions. Importantly, it is crucial to recognize the functions that microorganisms play in this adaptation. Soil bacteria belonging to the genus *Agrobacterium* are one type of microorganism that has a significant impact on the variety of plant life on Earth. In the field of biotechnology, *Agrobacterium tumefaciens* is a plant pathogenic bacterium that can cause Crown Gall disease by transferring a segment of oncogenic DNA, also known as T DNA (transfer DNA), to susceptible plant cells on a large plasmid known as the tumor inducing or Ti plasmid. The current article primarily focuses on the relatively underappreciated role of *Agrobacterium* in providing much-needed plant diversity to aid in the preservation of ecological equilibrium, which benefits society, and examines this role based on literature investigations. We begin by outlining the role that underlying molecular and cellular mechanisms play in the mutualistic relationship between *Agrobacterium* and plant species, which controls plant biodiversity. Next, we look at the crucial role that *agrobacterium* has played in the adaptability and evolution of the plant kingdom. We have finally illuminated the critical function *Agrobacterium* plays in the conservation of biodiversity. We end by warning that, despite the fact that *Agrobacterium* has been crucial to the development of biotechnology, there are a lot of challenging problems related to its unintended impacts on biodiversity. A balanced strategy that considers the possible ecological consequences of genetic engineering procedures based on *Agrobacterium* is required to address these problems. Prioritizing sustainable approaches that reduce the risks associated with using *Agrobacterium* is essential as technology advances and ensures the long-term conservation of biodiversity.

Introduction

Generally speaking, biodiversity is the variety of life on Earth as a result of the intricate and interwoven network of species, ecosystems, and genetic variation [1]. Biodiversity means “the variety and variability of biological organisms” according to researchers working in fields that are focused on biodiversity, such as population biology, ecology, systematics, evolution, and genetics [1]. “Variability among living organisms from all sources” is the definition of biodiversity given by the Convention on Biological Diversity [2]. Even within species, there is genetic variation. Genetic recombination or mutation, genetic polymorphism (the existence of several variants of the same gene), isolation of gene pools, local selection pressures, and habitat (environmental) complexity are a few of the factors that can cause genetic variety within a species, which helps in adaptation in the environment [3,4]. For instance, due to genetic variety that allows the kinds to adapt to changing environmental conditions, numerous crop varieties survive [5].

Varieties with little to no genetic variation, however, might become vulnerable to biotic and abiotic stressors [6]. More importantly, even though large animals like tigers

and trees are typically associated with biodiversity preservation, microorganisms also have significant roles to play [7,8]. One such microbe is the genus *Agrobacterium* of soil bacteria, which has a significant influence on the variety of plant life on Earth [9-11]. The biodiversity of plants provides people with a wealth of benefits, including resilience to climate change and other perturbations and is the basis for our current food supply, which also includes functional food and medicine derived from plants [12,13]. The current paper thus primarily focuses on the relatively underappreciated role of *Agrobacterium* in providing much-needed plant diversity to aid in the preservation of ecological equilibrium, which benefits society, and examines this role based on literature investigations.

***Agrobacterium*: A natural genetic engineer**

In the realm of biotechnology, *Agrobacterium* is well known for its ability to introduce the Ti (tumor-inducing) plasmid—a segment of its DNA—into plant cells [14,15]. More specifically, *Agrobacterium tumefaciens* is a plant pathogenic bacterium, with the ability to deliver a segment of oncogenic DNA carried on a large plasmid called the tumor inducing or Ti plasmid to susceptible plant cells. On the other hand, nonpathogenic strains of *Agrobacterium* either lack these plasmids entirely or carry mutant forms of plasmids [14]. A strict requirement of the Ti plasmid for virulence in plants is further established by mutational analyses and the demonstration that Ti plasmids introduced into *Phyllobacterium spp.* found in soil or *Rhizobium*, a nitrogen-fixing organism, transform these nonpathogenic species into tumor-inducing pathogens in plants (a typical example is Crown Gall disease) [15,16]. Moreover, the *A. tumefaciens*-plant interactions are among the first where such host-released signals such as opines have been detected, since they are the signal transduction cascade that leads to the expression of virulence (*vir*) genes in response to these signals [17,18]. Notable in this context is the fact that the plant's metabolites, sugars, amino acids, and α -keto acids (collectively referred to as opines) are transformed into a range of low molecular weight compounds by the enzymes known as opine synthase, which are encoded with Ti-DNA and found in *Agrobacterium* [19,20]. These metabolites are then used by the *Agrobacterium* to continue growing. Two of the *vir* genes, *virA* and *virG*, are required for *A. tumefaciens* to respond to inducing conditions evoked by the plant, according to additional genetic study [21,22]. This implies that *A. tumefaciens* employs a clever two-component signal transduction mechanism, which is typically responsible for triggering complex responses to a variety of inputs, including hormonal regulation and environmental cues [23]. Moreover, Ti plasmid is generally transferred into plants by *Agrobacterium* using the Type IV Secretion System (T4SS), which is a biological "molecular syringe" produced by many gram-negative bacteria that transports proteins or DNA-protein complexes into other cells—a necessary step for horizontal gene transfer—with the aid of the two-component regulatory system that uses *virA* and *virG* [24]. *Agrobacterium* also employs quorum sensing, a method of cell-to-cell communication that enables bacteria to exchange information about cell density and modify gene expression appropriately, in addition to T4SS and other systems like Type VI secretion systems, etc [25,26]. This allows for the conjugative

transfer of the Ti plasmid between bacteria during plant pathogenesis when specific opines (inducing factors) derived from plants are present. Because of this special quality, *Agrobacterium* is useful in genetic engineering as it allows specific genes to be inserted into plants [27]. However, because of its enormous impact on biodiversity in natural ecosystems, its significance extends beyond the lab.

Plant biodiversity is controlled by the mutualistic relationship between *agrobacterium* and plant species. Numerous plant species and *Agrobacterium* have mutualistic relationships [28,29]. The plant receives genes that control its growth, development, and response to environmental stressors from *Agrobacterium* in exchange for nutrients and a safe habitat within plant tissues. The biodiversity of plants may be greatly impacted by this interaction. To elucidate, the ability of *Agrobacterium* to transfer DNA into plants for stable genetic transformation or transient expression may underpin plant diversification [30]. This allows plants to develop altered phenotypic features in response to a range of environmental stimuli. Nevertheless, even though the process of T-DNA transfer and integration has been thoroughly investigated since the early 1970s, many facets of the underlying mechanism remain unclear to date. For example, little is known about the process by which T-DNA integrates into the genome of the host [plant]. It is tempting to speculate that the variety in T-DNA integration into a particular plant genome locus may play a key role in modifying a wide range of phenotypic features in plants [31,32].

Some early research appears to indicate that integrated T-DNAs are preferentially located in transcriptionally active chromatin [33,34]. However, these investigations relied on transgenic plant analyses that were regenerated under antibiotic selection using a reporter genetic fragment that was resistant to specific antibiotics and was engineered into T-DNA [11]. Because of this, it was nearly difficult to find evidence of integration into heterochromatin, which does not support the transcription of the antibiotic-resistant reporter. Indeed, an entirely different result was observed in experiments carried out without the use of antibiotics, showing that T-DNA integrated randomly in every region of chromatin, even though specific epigenetic markers might control certain loci integration—the epigenetic signature of which may, among other things, govern the biodiversity of plants [28,35].

Moreover, it has been proposed that nucleosome histones, such as H2A, assist in the targeting of the T-DNA complex to the host chromatin upon its nuclear entry by permitting interaction between the T-DNA complex and the host chromatin prior to integration as *Agrobacterium* does not encode a dedicated integrase among its effector proteins, unlike many integrating viruses [28]. Thus, it is plausible that the class of histone proteins may control where T-DNA integration occurs within the plant genome. This could modify the plant's gene expression pattern and enable the plant to grow and survive in a particular environment with an altered phenotype as a consequence of a mutualistic relationship between *Agrobacterium* and particular plant species [36]. However, to support this assumption made based on early evidence found in the literature, more investigation is required. Alternatively, since

T-DNA is transferred to plant cells and integrates into the plant genome, it would appear that Ti plasmids can be used as a vector to introduce foreign genes into plant cells, thereby enhancing the biodiversity of the plant kingdom. A substantial body of literature describes the procedures for introducing foreign genes into plants using *Agrobacterium*, although it is outside the scope of this theme article.

Agrobacterium evades plant immunological defense system prior to the transfer of T-DNA into host cells. When microorganisms like *Agrobacterium* try to invade their hosts, they typically encounter immunological resistance because host cells have what are known as “microbe- or Pathogen-Associated Molecular Patterns” (MAMPs or PAMPs) [37,38]. These so-called “non-self” molecules start PAMP-Triggered Immunity (PTI), the initial defense mechanism. In response, pathogens try to neutralize PTI activation by introducing specific effector proteins into the cytoplasm of their host [39,40]. When cognate intracellular plant proteins perceive these pathogen-encoded effectors, Effector-Triggered Immunity (ETI), the second line of defense, is activated [41,42]. Thus, in addition to limiting the spread of pathogens, plants with the ability to activate ETI can also strengthen themselves against future attacks.

Flagellin and EF-Tu are the two oligopeptides that are recognized by FLS2 and EFR, the two MAMP receptors in plants that have been studied the most [43]. These membrane-located Leucine-Rich Repeat-Receptor-Like Kinases (LRR-RLK) are able to transform and convey perceived “attack signals” into the inside of cells in order to trigger the proper defense responses because of their composite structure. On the other hand, the primary goals of pathogens like *Agrobacterium* are to take advantage of their hosts’ nutrition and proliferate to a high degree. In plants, microbial PAMP recognition by plant PRRs is facilitated by Mitogen-Activated Protein Kinase (MAPK) cascades [44]. The prompt activation of genes encoding effector proteins and transcription factors in response to the pathogen’s invasion is caused by this recognition.

In essence, by avoiding plant immunity and incorporating T-DNA into the plant genome, *Agrobacterium* have collectively contributed to the spread of genetic variety throughout plant populations and the evolution of plants. Plants can adapt to shifting environmental conditions because of their diversity, which serves as the foundation for evolution. New traits and attributes that give a certain plant an advantage in its natural ecosystem may result from it. As a result, new plant species may eventually appear, adding to the biodiversity.

Plant evolution and adaptation: the function of *agrobacterium*

The movement of genetic information across organisms, such as bacteria and plants, is known as Horizontal Gene Transfer (HGT) [45]. This process includes the dissemination of antibiotic resistance genes among bacteria (apart from those passed from parent to offspring), which promotes the evolution of pathogens. Bacteria and archaea are part of the microbial world, where HGT is a common occurrence. To stress this point even more, HGT mechanisms are well-characterized and thought to be essential to

the evolution of species in this field [46]. For example, there has been evidence in *Nicotiana*, *Linaria*, and, more recently, *Ipomoea* species, of T-DNAs from *Agrobacterium* spp. entering plant genomes and being maintained in the germline [47]. Crucially, the genes that were transferred seem to have played a part in the evolution of these plants because they do not cause the typical disease phenotype in the plant.

The impact of T-DNA acquisition on evolution, in theory primarily depends on a gene’s ability to be stably integrated into the genome of a plant after transfer; the recipient plant genome containing the T-DNA must remain intact following genomic rearrangements during subsequent cell division; the transformed cell must enter the germline; and the integrated sequence must be preserved over the course of evolution [31,48]. The latter is most likely to happen if the gene confers a selective advantage on the recipient organism, enabling it to thrive in a hostile environment. But one would envision an additional specific requirement, particularly in the case of T-DNA genes integration into plant genomes, which is that the inserted genes must somehow be modified or regulated from their “natural” expression pattern which allows for vigorous cell growth, the long-term expression of which would be detrimental to the plant’s survival [49,50].

As of now, it is unclear how plants have managed to evade the *Agrobacterium*-programmed expression of T-DNA sequences upon insertion into their genomes, however a couple of theories may be taken into consideration. First, it’s often thought that the T-DNA may have integrated in a heterochromatin region—a transcriptionally inactive region of the genome—which is a trait that the inserted T-DNA then acquires to inhibit the expression of its genes [51,52]. Second, accumulating data appear to indicate the presence of foreign IbT-DNAs in sweet potatoes, which were initially discovered using short RNA sequencing and assembly. After being introduced into the plant genome as an inverted repeat, IbT-DNA is silenced even after it has integrated into a transcriptionally active DNA region. This is accomplished by the plant RNA silencing complex acting upon the IbT-DNA with a trans-acting effect [53,54].

The conservation of biodiversity: the role of *agrobacterium*

Agrobacterium, primarily known for its use in genetic engineering, also plays a crucial role in the conservation of biodiversity [55,56]. Its unique ability to transfer genes into plants has found application in preserving endangered species, restoring ecosystems, and enhancing agricultural sustainability. Therefore, it is essential for conservation efforts to comprehend how *Agrobacterium* contributes to plant diversity. The genetic variety and adaptability of plant species are also protected when we maintain natural habitats in which *Agrobacterium* interacts with plants. This is essential in light of environmental issues such as climate change. Essentially, *Agrobacterium* plays a multifaceted role in the preservation of biodiversity [57]. First, it helps to preserve genetic diversity. *Agrobacterium*-mediated transformation allows for the introduction of desirable traits into plants, aiding in the conservation of genetic diversity. This is particularly significant for

endangered species with limited populations, as it can help address challenges such as inbreeding depression and susceptibility to diseases [58]. Therefore, understanding the genetic composition and structure of these species in their natural habitat is essential to developing viable management methods for the protection of rare and endangered plant species.

Second, since habitat loss and changing environmental circumstances are common hazards to endangered plant species, *Agrobacterium* plays a crucial role in ecosystem restoration efforts [59,60]. *Agrobacterium* provides a tool for introducing genes that confer resistance to particular conditions, thereby offering a way to protect endangered species in their native environments [61]. Genetic variation is considered necessary for plant species to survive in their natural habitat, as the survival of these species is believed to depend on a range of environmental factors that affect the number of alleles on different loci in the genome [62,63]. Third, endangered plant species often face threats like habitat loss and changing environmental conditions. *Agrobacterium* offers a tool to insert genes that confer resistance to specific stresses, providing a means to safeguard rare species in their natural habitats [61,62,64]. Fourth, *Agrobacterium* supports sustainable agriculture in addition to conservation.

Increased resistance to pests and diseases in modified crops lowers the requirement for chemical inputs, supporting eco-friendly farming methods and preserving the biodiversity of agroecosystems [65,66]. Fifth, *Agrobacterium* is also involved in cryopreservation techniques, where plant tissues or seeds are stored at extremely low temperatures [67]. This method helps preserve the genetic material of endangered plants, serving as a backup for biodiversity conservation efforts. Finally, understanding the role of *Agrobacterium* in plant-microbe interactions is essential for maintaining healthy soils. As a natural soil bacterium, *Agrobacterium* influences plant growth and nutrient cycling, indirectly contributing to the overall biodiversity of terrestrial ecosystems [68]. Despite its potential benefits, the use of *Agrobacterium* in conservation raises ethical concerns. Striking a balance between preserving biodiversity and potential ecological risks associated with genetic modifications requires careful consideration and adherence to strict ethical guidelines [69,70].

Conclusion & Challenges

In conclusion, *Agrobacterium's* role in biodiversity conservation extends beyond biotechnology labs. Its applications in genetic engineering contribute to the protection of endangered species, ecosystem restoration, and sustainable agriculture. However, responsible and ethical use is imperative to ensure that these interventions do not inadvertently harm the very biodiversity they seek to preserve. Essentially, *Agrobacterium* has evolved from a phytopathogen to a potent transgenic vector thanks to its remarkable ability to transfer its genetic material to host cells. For instance, for crop breeding and molecular plant biology research, stable transformation is the most extensively utilized technique for producing transgenic plants. Over time, *Agrobacterium*-mediated HGT has advanced significantly, providing a fuller understanding of

the diversity within the kingdom of plants. However, the unintended consequences of its presence pose significant challenges to the conservation of biodiversity. For example, *Agrobacterium*-mediated genetic transformation can lead to unintended gene flow and hybridization between Genetically Modified Organisms (GMOs) and their wild counterparts [71].

This could lead to the extinction of native species and a reduction in biodiversity by endangering the genetic integrity of natural plant populations [72]. Moreover, the dynamics of natural ecosystems may be upset by the introduction of genetically engineered plants containing genes generated from *Agrobacterium*. These alterations could have an impact on how various species interact with one another, which could have a domino effect on the food web and affect the variety and abundance of other organisms in the environment [73,74]. Additionally, the use of *Agrobacterium* in biotechnology can inadvertently affect non-target organisms, including beneficial insects, soil microbes, and other wildlife [75]. Alterations in trophic relationships, alterations in soil microbial populations, and disturbances in pollination patterns are some of the possible ecological repercussions that could have a negative impact on biodiversity. Weeds resistant to *Agrobacterium* may evolve as a result of genetic engineering procedures that expose the organism to continuous exposure.

These resistant plants have the potential to outcompete native flora, increasing the difficulties brought on by invasive species and endangering the variety of natural ecosystems. Also, unexpected ecological repercussions could arise from the release of genetically modified organisms into the environment that carry genes produced from *Agrobacterium*. These repercussions may include adjustments to the ecological roles of certain plant species, variations to the nutrient cycling, and shifts in the interactions between pollinators and plants, all of which can have an adverse effect on biodiversity. Taken together, even while *Agrobacterium* has been essential in the advancement of biotechnology, there are a number of difficult issues associated with its unintentional effects on biodiversity. A balanced approach to resolving these concerns is necessary, taking into account the potential ecological implications of *Agrobacterium*-based genetic engineering processes. As technology continues to evolve, it is imperative to prioritize sustainable practices that minimize the risks associated with the use of *Agrobacterium*, ensuring the long-term conservation of biodiversity.

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References

1. National Research Council (US) Committee on noneconomic and economic value of biodiversity. Perspectives on biodiversity (1999) Valuing its role in an ever-changing world. Washington (DC): National Academies Press, USA.
2. Naeem S, Chazdon R, Duffy JE, Prager C, Worm B (1844) Biodiversity and human well-being: an essential link for sustainable development. Proc Biol Sci 283(1844): 20162091.

3. Saeb AT, Naqeb D (2016) The impact of evolutionary driving forces on human complex diseases: A population genetics approach. *Scientifica (Cairo)* 16: 2079704.
4. Lee CR, Mitchell Olds T (2012) Environmental adaptation contributes to gene polymorphism across the *Arabidopsis thaliana* genome. *Mol Biol Evol* 29(12): 3721-3728.
5. Salgotra RK, Chauhan BS (2023) Genetic diversity, conservation, and utilization of plant genetic resources. *Genes (Basel)* 14(1): 174.
6. Paliwal S, Tripathi MK, Tiwari S, Tripathi N, Payasi DK, et al. (2023) Molecular advances to combat different biotic and abiotic stresses in linseed (*Linum usitatissimum* L): A comprehensive review. *Genes (Basel)* 14(7): 1461.
7. Nulkar G, Bedarkar M, Ghate K, Nulkar S (2021) Hitchhiking microbes: Declining biodiversity & emerging zoonoses. *Indian J Med Res* 153(3): 367-374.
8. Vero L, Boniotti MB, Budroni M, Buzzini P, Cassanelli S, et al. (2019) Preservation, characterization and exploitation of microbial biodiversity: The perspective of the Italian network of culture collections. *Microorganisms* 7(12): 685.
9. Gupta A, Gupta R, Singh RL (2016) Microbes and environment. Principles and applications of environmental biotechnology for a sustainable future 15: 43-84.
10. Souza Rd, Ambrosini A, Passaglia LM (2015) Plant growth-promoting bacteria as inoculants in agricultural soils. *Genet Mol Biol* 38(4): 401-419.
11. Gelvin SB (2003) *Agrobacterium*-mediated plant transformation: the biology behind the "gene-jockeying" tool. *Microbiol Mol Biol Rev* 67(1): 16-37.
12. Alves RR, Rosa IM (2007) Biodiversity, traditional medicine and public health: where do they meet? *J Ethnobiol Ethnomed* 3: 14.
13. Sen T, Samanta SK (2015) Medicinal plants, human health and biodiversity: a broad review. *Adv Biochem Eng Biotechnol* 147: 59-110.
14. Gordon JE, Christie PJ (2014) The *Agrobacterium* Ti Plasmids. *Microbiol Spectr* 2(6): 10.
15. Hwang HH, Yu M, Lai EM (2017) *Agrobacterium*-mediated plant transformation: biology and applications. *Arabidopsis Book* 15: e0186.
16. Moënne LY, Mavingui P, Combes C, Normand P, Steinberg C (2014) Microorganisms and biotic interactions. *Environmental microbiology: Fundamentals and applications* 29: 395-444.
17. Wang C, Ye F, Chang C, Liu X, Wang J, et al. (2019) *Agrobacteria* reprogram virulence gene expression by controlled release of host-conjugated signals. *Proc Natl Acad Sci USA* 116(44): 22331-22340.
18. Brencic A, Winans SC (2005) Detection of and response to signals involved in host-microbe interactions by plant-associated bacteria. *Microbiol Mol Biol Rev* 69(1): 155-194.
19. McFarlane JS, Davis CL, Lamb AL (2018) Staphylopin, Pseudopaline, and Yersinopine dehydrogenases: A structural and kinetic analysis of a new functional class of opine dehydrogenase. *J Biol Chem* 293(21): 8009-8019.
20. Flores AL, Eberhard A, Winans SC (2012) *Agrobacterium tumefaciens* can obtain sulphur from an opine that is synthesized by octopine synthase using S-methylmethionine as a substrate. *Mol Microbiol* 84(5): 845-856.
21. Brown PJB, Chang JH, Fuqua C (2023) *Agrobacterium tumefaciens*: a transformative agent for fundamental insights into host-microbe interactions, Genome Biology, Chemical Signaling, and Cell Biology. *J Bacteriol* 205(4): e0000523.
22. Stachel SE, Zambryski PC (1986) virA and virG control the plant-induced activation of the T-DNA transfer process of *A. tumefaciens*. *Cell* 46(3): 325-333.
23. Mukhopadhyay A, Gao R, Lynn DG (2004) Integrating input from multiple signals: the VirA/VirG two-component system of *Agrobacterium tumefaciens*. *Chembiochem* 5(11): 1535-1542.
24. Li YG, Christie PJ (2018) The *Agrobacterium* VirB/VirD4 T4SS: Mechanism and architecture defined through *In Vivo* mutagenesis and chimeric systems. *Curr Top Microbiol Immunol* 418: 233-260.
25. Lang J, Faure D (2014) Functions and regulation of quorum-sensing in *Agrobacterium tumefaciens*. *Front Plant Sci* 5: 14.
26. Wu CF, Smith DA, Lai EM, Chang JH (2018) The *Agrobacterium* type VI secretion system: A contractile nanomachine for interbacterial competition. *Curr Top Microbiol Immunol* 418: 215-231.
27. Nester EW (2015) *Agrobacterium*: nature's genetic engineer. *Front Plant Sci* 5: 730.
28. Lacroix B, Citovsky V (2019) Pathways of DNA transfer to plants from *Agrobacterium tumefaciens* and related bacterial species. *Annu Rev Phytopathol* 57: 231-251.
29. Gelvin SB (2009) *Agrobacterium* in the genomics age. *Plant Physiol* 150(4): 1665-1676.
30. Lacroix B, Citovsky V (2016) A functional bacterium-to-plant DNA transfer machinery of rhizobium etli. *PLoS Pathog* 12(3): e1005502.
31. Gelvin SB (2021) Plant DNA repair and *Agrobacterium* T-DNA Integration. *Int J Mol Sci* 22(16): 8458.
32. Gelvin SB (2017) Integration of *Agrobacterium* T-DNA into the Plant Genome. *Annu Rev Genet* 51: 195-217.
33. Shilo S, Tripathi P, Melamed BC, Tzfadia O, Muth TR, et al. (2017) T-DNA-genome junctions form early after infection and are influenced by the chromatin state of the host genome. *PLoS Genet* 13(7): e1006875.
34. Hsieh JA, Chang P, Kuang LY, Hsing YC, Chen PY (2023) Rice transformation treatments leave specific epigenome changes beyond tissue culture. *Plant Physiol* 193(2): 1297-1312.
35. Gelvin SB, Kim SI (2007) Effect of chromatin upon *Agrobacterium* T-DNA integration and transgene expression. *Biochim Biophys Acta* 1769(5-6): 410-21.
36. Kim JH, Castroverde CDM (2020) Diversity, function and regulation of cell surface and intracellular immune receptors in *Solanaceae*. *Plants (Basel)* 9(4): 434.
37. Barton IS, Fuqua C, Platt TG (2018) Ecological and evolutionary dynamics of a model facultative pathogen: *Agrobacterium* and crown gall disease of plants. *Environ Microbiol* 20(1): 16-29.
38. Thieffry A, López MD, Bornholdt J, Malekroudi MG, Bressendorff S, et al. (2022) PAMP-triggered genetic reprogramming involves widespread alternative transcription initiation and an immediate transcription factor wave. *Plant Cell* 34(7): 2615-2637.
39. Pitzschke A (2013) *Agrobacterium* infection and plant defense-transformation success hangs by a thread. *Front Plant Sci* 4: 519.
40. Zhang S, Li C, Si J, Han Z, Chen D (2022) Action mechanisms of effectors in plant-pathogen interaction. *Int J Mol Sci* 23(12): 6758.
41. Hatsugai N, Igarashi D, Mase K, Lu Y, Tsuda Y, et al. (2017) A plant effector-triggered immunity signaling sector is inhibited by pattern-triggered immunity. *EMBO J* 36(18): 2758-2769.
42. Newman MA, Sundelin T, Nielsen JT, Erbs G (2013) MAMP (Microbe-Associated Molecular Pattern) triggered immunity in plants. *Front Plant Sci* 4: 139.s
43. Lee CC, Wu YJ, Hsueh CH, Huang YT, Hsu YH (2018) Mitogen-activated protein kinase phosphatase 1 reduces the replication efficiency of Bamboo mosaic virus in *Nicotiana benthamiana*. *Mol Plant Pathol* 19(10): 2319-2332.

44. Emamalipour M, Seidi K, Zununi S, Jahanban A, Jaymand M, et al. (2020) Horizontal gene transfer: From evolutionary flexibility to disease progression. *Front Cell Dev Biol* 8: 229.
45. Tiwari P, Bae H (2020) Horizontal gene transfer and endophytes: An implication for the acquisition of novel traits. *Plants (Basel)* 9(3): 305.
46. Huamanquispe DG, Gheysen G, Kreuze JF (2015) Horizontal gene transfer contributes to plant evolution: The case of *Agrobacterium* T-DNAs. *Front Plant Sci* 24: 8.
47. Pikaard CS, Mittelsten Scheid O (2014) Epigenetic regulation in plants. *Cold Spring Harb Perspect Biol* 6(12): a019315.
48. Sedeek KEM, Mahas A, Mahfouz M (2019) Plant genome engineering for targeted improvement of crop traits. *Front Plant Sci* 10: 114.
49. Magori S, Citovsky V (2011) Epigenetic control of *Agrobacterium* T-DNA integration. *Biochim Biophys Acta* 1809(8): 388-3894.
50. Jupe F, Rivkin AC, Michael TP, Zander M, Motley ST, et al. (2019) The complex architecture and epigenomic impact of plant T-DNA insertions. *PLoS Genet* 15(1): e1007819.
51. Kyndt T, Quispe D, Zhai H, Jarret R, Ghislain M, et al. (2015) The genome of cultivated sweet potato contains *Agrobacterium* T-DNAs with expressed genes: An example of a naturally transgenic food crop. *Proc Natl Acad Sci U S A* 112(18): 5844-5849.
52. Khan S, Qurainy F, Nadeem M (2012) Biotechnological approaches for conservation and improvement of rare and endangered plants of Saudi Arabia. *Saudi J Biol Sci* 19(1): 1-11.
53. Supple MA, Shapiro B (2018) Conservation of biodiversity in the genomics era. *Genome Biol* 19(1): 131.
54. Pandit MA, Kumar J, Gulati S, Bhandari N, Mehta P, et al. (2022) Major biological control strategies for plant pathogens. *Pathogens* 11(2): 273.
55. Rahman SU, Khan MO, Ullah R, Ahmad F, Raza G (2023) *Agrobacterium*-mediated transformation for the development of transgenic crops; present and future prospects. *Mol Biotechnol*.
56. Heywood VH (2019) Recovering threatened plant species and their habitats: The need for integrated action. *Plant Divers* 41(2): 33-35.
57. Kim S, Park HJ, Lee CW, Kim NY, Hwang JE, et al. (2022) Endangered plant species under differing anthropogenic interventions: how to preserve *Pterygopleurum neurophyllum* in Wondong wetland? *PeerJ* 10: e14050.
58. Tiwari M, Mishra AK, Chakrabarty D (2022) *Agrobacterium*-mediated gene transfer: recent advancements and layered immunity in plants. *Planta* 256(2): 37.
59. Anderson JT, Willis JH, Mitchell T (2011) Evolutionary genetics of plant adaptation. *Trends Genet* 27(7): 258-66.
60. Szczecińska M, Sramko G, Wołosz K, Sawicki J (2016) Genetic diversity and population structure of the rare and endangered plant species *Pulsatilla patens* (L.) mill in east central Europe. *PLoS One* 11(3): e0151730.
61. Ahanger MA, Akram NA, Ashraf M, Alyemeni MN, Wijaya L, et al. (2017) Plant responses to environmental stresses-from gene to biotechnology. *AoB Plants* 9(4): plx025.
62. Massa F, Defez R, Bianco C (2022) Exploitation of plant growth promoting bacteria for sustainable agriculture: Hierarchical approach to link laboratory and field experiments. *Microorganisms* 10(5): 865.
63. Pinski A, Betekhtin A (2023) Efficient *Agrobacterium*-mediated transformation and genome editing of *Fagopyrum tataricum*. *Front Plant Sci* 14: 1270150.
64. Blanc G, Baptiste C, Oliver G, Martin F, Montoro P (2006) Efficient *Agrobacterium tumefaciens*-mediated transformation of *embryogenic calli* and regeneration of *Hevea brasiliensis* Müll Arg. plants. *Plant Cell Rep* 24(12): 724-733.
65. Nadarajah K, Abdul NS (2021) Plant-microbe interaction: Aboveground to belowground, from the good to the bad. *Int J Mol Sci* 22(19): 10388.
66. Ormandy EH, Dale J, Griffin G (2011) Genetic engineering of animals: ethical issues, including welfare concerns. *Can Vet J* 52(5): 544-550.
67. Bawa AS, Anilakumar KR (2013) Genetically modified foods: safety, risks and public concerns-a review. *J Food Sci Technol* 50(6): 1035-1046.
68. Buiatti M, Christou P, Pastore G (2013) The application of GMOs in agriculture and in food production for a better nutrition: two different scientific points of view. *Genes Nutr* 8(3): 255-270.
69. Wilson EO, Peter FM (1998) The loss of diversity causes and consequences. *Biodiversity*. National Academies Press (US), Washington (DC), USA.
70. Ream W (2009) Genetically engineered plants: greener than you think. *Microb Biotechnol* 2(4): 401-405.
71. National Research Council (US) Committee on Environmental Impacts Associated with Commercialization of Transgenic Plants. *Environmental Effects of Transgenic Plants: The Scope and Adequacy of Regulation*. Washington (DC): National Academies Press (US); 2002. 1, Ecological, Genetic, and Social Factors Affecting Environmental Assessment of Transgenic Plants.
72. Ladics GS, Bartholomaeus A, Bregitzer P, Doerrner NG, Gray A, et al. (2015) Genetic basis and detection of unintended effects in genetically modified crop plants. *Transgenic Res* 24(4): 587-603.
73. Rodríguez A, Rangseekaew P, Lasudee K, Pathom W, Manzanera M (2021) Impacts of agriculture on the environment and soil microbial biodiversity. *Plants (Basel)* 10(11): 2325.
74. Schütte G, Eckerstorfer M, Rastelli V, Reichenbecher W, Restrepo S, et al. (2017) Herbicide resistance and biodiversity: agronomic and environmental aspects of genetically modified herbicide-resistant plants. *Environ Sci Eur* 29(1): 5.
75. National Research Council (US) (2004) Committee on Identifying and Assessing Unintended Effects of Genetically Engineered Foods on Human Health. *Safety of Genetically Engineered Foods: Approaches to Assessing Unintended Health Effects*. Washington (DC): National Academies Press (US), Unintended Effects from Breeding.