



The Evaluation of the Speed-Oligo® Mycobacteria Assay for Identification of *Mycobacterium* spp. from Smear Positive and Negative Sputum Samples

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Abstract

Speed-oligo® Mycobacteria is an oligochromatographic test for the qualitative detection of *Mycobacterium* genus and the species of *M. tuberculosis* complex, directly in clinical samples. It is based on polymerase chain reaction targeting 16S rRNA and 16S-23S rRNA regions and double-reverse hybridization on a dipstick using probes bound to colloidal gold and to the membrane. In this study, we evaluated 25 type reference strains of mycobacteria (18 non-tuberculosis mycobacteria, *M. tuberculosis* H37Rv and *M. tuberculosis* H37Ra), 60 sputum samples (40 smear positive, 20 smear negative) collected from patient with suspected TB. All results were compared with microscopy, Löwenstein Jensen culture and Inno-Lipa (GenoType *Mycobacterium* CM/AS; Hain Lifescience, Germany). All smear positive sputum samples were positive with microscopy, culture and Speed-oligo® Mycobacteria. Of 20 smear negative sputum samples, 7 were culture positive. Of 7 culture positive samples, 3 were positive with microscopy and Speed-oligo® Mycobacteria. It is not effective to identify for *M. intermedium*, *M. kansasii* and *M. xenopi*.

Keywords: Tuberculosis; NTM; Speed oligo; PCR

Introduction

Tuberculosis (TB) is a chronic infectious disease that is common all over the World [1]. In recent years, nontuberculous mycobacteria (NTM) infections are getting increase depend on a immunosuppression, AIDS, malignancy, drug abuse, bad living conditions or medical intervention. NTM causing clinical disease have become increasingly frequent and more varied. The incidence and prevalence of NTM lung disease are increasing worldwide [2-4]. The rapid differentiation between NTM and *M. tuberculosis* complex (MTBC) is primary importance for effective and accurate treatment protocol [5]. Mycobacteriology laboratory findings have an important role in the diagnosis and control of this disease. MTBC and NTM can not be distinguished by microscopy. Diagnosis with culture method is time consuming. Conventional identification tests at the species level are quite complex and sometimes no definite conclusions can be drawn. PCR based methods are widely used in routine diagnostic laboratories for rapid diagnosis [5-8]. Speed-oligo® Direct *Mycobacterium* tuberculosis (SO-DMT) is an oligochromatographic test (Vircell SL, Santa Fe, Granada, Spain) for the specific identification of *M. tuberculosis* complex (MTC) from *Mycobacterium* genus in respiratory specimens. It is based on

polymerase chain reaction targeting 16S rRNA and 16S-23S rRNA regions and double-reverse hybridization on a dipstick using probes bound to colloidal gold and to the membrane. It may represent a fast and easy alternative for differentiating between MTBC and NTM in direct samples at laboratories with standard laboratory equipment (thermocycler and thermoblock) [5-8]. The aim of this study was to evaluate SO-DMT by using 25 type reference strains of mycobacteria (18 non-tuberculosis mycobacteria, *M. tuberculosis* H37Rv and *M. tuberculosis* H37Ra), 60 sputum samples (40 smear positive, 20 smear negative) collected from patient with suspected TB.

Material and Methods

Specimen group

In this study, we evaluated 20 type reference strains of mycobacteria (18 non-tuberculosis mycobacteria, *M. tuberculosis* H37Rv and *M. tuberculosis* H37Ra), 60 sputum samples (40 smear positive, 20 smear negative) collected from patient with suspected TB.

Specimen processing, microscopy, culture, and storage conditions

Respiratory specimens were decontaminated with 4% NaOH-NALC method, directly examined (with Ehrlich-Ziehl-Neelsen staining method), and cultured by Löwentein Jensen medium and *Mycobacterium* Indicator Growth System (Becton Dickinson, USA). *Mycobacteria* were identified by the Geno Type *Mycobacterium* CM/AS assay (Hain Lifescience, Nehren, Germany). At least one aliquot of all decontaminated specimens was maintained for a maximum of 7 days at 4 °C until assayed with SO-DMT; if the assay procedure was delayed, samples were frozen at -80 °C until their analysis [9-13].

SO-DMT assay

The SO-DMT assay was carried out as manufactures instructions. Positive and negative controls were used in each test run. The SO-DMT comprises four steps: DNA extraction, amplification, PCR

product detection, and result reading, as detailed below. DNA is extracted from a previously concentrated decontaminated sample using the reagents included in the kit according to the manufacturer's instructions. Invalid assay results (absence of the human amplification control line) are usually obtained from specimens with a volume of <200µl. Sample preparation of a set of 12 specimens takes approximately 45min. DNA amplification is performed with 10µl of disrupted sample supernatant and 15µl of the ready-to-use PCR mix. The amplification process is a multiplex PCR in which a triple target is amplified. DNA extraction is controlled by amplification of the human gene RNase P, which is also used to control for the presence of PCR inhibitors. The presence of *Mycobacterium* is assayed by amplifying a fragment of the 16S rRNA sequence and the presence of MTBC members by amplifying a fragment of the insertion sequence IS6110. DNA amplification was performed using a standard thermocycler and took 1h to complete. A dipstick with probes bound to colloidal gold and to the membrane is used for PCR product detection (Figure 1).

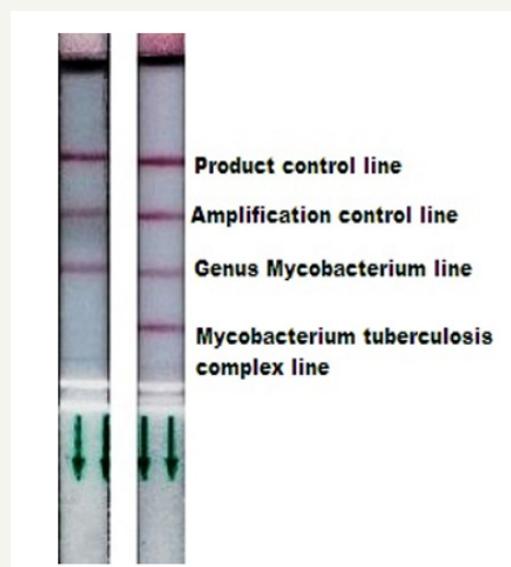


Figure 1: The PCR product is denatured for 2min and added to a preheated running solution in a thermoblock. After introduction of the dipstick and a 5min incubation period, the result is obtained from the presence of visible lines by using an automatic reading system. In our study, the results interpretation was performed automatically by the software according to the intensity of the line, but a visual interpretation is also possible based on the presence or absence of the red color on control and test lines.

The dipstick has four lines: a product control line, a PCR amplification control line, a test line for the whole *Mycobacterium* genus, whose presence in the absence of the *M. tuberculosis* complex line indicates the presence of NTM genetic material, and a test line for MTC members [14].

Results

All results were compared with Löwenstein Jensen culture. All smear positive sputum samples were positive with culture and

Table 1: The positivity rate of the SO-DMT in smear positive and smear negative sputum samples when culture results were taken as gold standards.

Specimens	SO-DMT assay			
	True Positive (%)	True Negative (%)	False Positive (%)	False Negative (%)
Smear positive(n:60)	60(100)	-	-	-
Smear negative(n:20)	3(15)	13(65)	-	4(20)

Speed-oligo® *Mycobacteria*. Of 20 smear negative sputum samples, 7 were culture positive. Of 7 culture positive samples, 3 were positive SO-DMT (Table 1). SO-DMT assay showed no reactivity in any of the mycobacterium-free specimens or in those with 18 NTM reference strains. When LJ culture results were accepted as a reference method, the sensitivity rate of SO-DMT was 100 % for smear positive specimens and 42% for smear positive specimens (Table 2).

Table 2: The sensitivity, specificity, PPV and NPV of SO-DMT assay in smear positive and smear negative sputum samples culture results were taken as gold standard.

Specimens	Sensitivity	Specificity	PPVA	NPVB
Smear positive(n:60)	100	-	100	-
Smear negative(n:20)	42	100	100	76

APositive predictive value, BNegative predictive value

Discussion

TB is still one of the most contagious and deadly infections with approximately a quarter of cases not being identified and/or treated mainly due to a lack of resources. In the definitive diagnosis of tuberculosis, smear staining from clinical specimens and culture is gold standard. However, the sensitivity of the microscopy test is low. Culture requires an incubation period of 2-8 weeks for definitive diagnosis [15-19]. In general, patients with negative sputum microscopy at the first examination are being treated until the end of culture. Due to the contamination of the tuberculosis bacillus by droplets, patients with negative sputum microscopy may become infected during this period. These uncertainties in the diagnosis and treatment of tuberculosis play an important role in spreading the disease and worsening the disease in the patient [1-8]. The incidence of tuberculosis is increasing all over the world. Due to the increasing prevalence of multiple germline resistant strains and the emergence of a variety of immunosuppressive diseases such as AIDS, the use of faster and more sensitive diagnostic methods in the early diagnosis of this disease has become compulsory [20-22].

For this purpose, PCR - based automated and semi-automated kit based PCR systems developed specifically for MTBC diagnosis are widely used in routine diagnostic laboratories. The sensitivity and specificity of these methods vary according to the system used [6]. In a various studies, lung-derived specimens were analyzed by FDA approved Cobas Amplicor MTB (CAMPMTB; Roche Diagnostic Systems, Inc., Branchburg, NJ) and Amplified *Mycobacterium tuberculosis* Direct Test (AMTDII; Gen-Probe, Inc., San Diego, Calif). The sensitivity of these tests were between 55% and 97% and the specificity between 92% and 100% [6,8,10]. In our study, 60 sputum samples (40 smear positive, 20 smear negative) in smear positive and negative sputum samples collected from patient with suspected TB, 20 type reference strains of mycobacteria (18 non-tuberculosis mycobacteria, *M. tuberculosis* H37Rv and *M. tuberculosis* H37Ra) were evaluated with SO-DMT to determine sensitivity and specificity. When LJ culture results were accepted as a reference method, the sensitivity rate of SO-DMT was 100 % for smear positive specimens and 42% for smear positive specimens. The sensitivity of the assay was very satisfactory in AFB smear-positive specimens and correctly classified samples as MTBC or NTM. The sensitivity was lower (42%) in AFB smear-negative samples within the range previously reported for smear-negative samples [6,22]. The overall specificity was very high (100%). Several commercially available nucleic acid amplification tests have been extensively evaluated for the detection of MTBC species in respiratory samples, showing high specificity and good positive predictive values for smear-positive samples and variable sensitivity for smear-negative and extrapulmonary TB samples [6].

However, most of these tests require the use of real-time PCR or precision instruments that are not affordable in many countries with a high TB burden, or they involve technologies that require extensive hands-on time. In contrast, the SO-DMT assay combines a PCR amplification method with a simple detection procedure that does not require long post amplification preparation or exclusive equipment, reducing manipulation to only two pipetting steps. As limitations, assay automation is not possible and separate laboratory areas should be used for specimen preparation, target amplification, and amplicon detection.

In conclusion, the SO-DMT assay requires only 30min of hands-on time and offers a total turnaround time of 2h from sample reception. This method directly detects *Mycobacterium* species in a clinical specimen and simultaneously differentiates them from MTBC species. Results obtained in a convenience sample of AFB smear-positive specimens suggest that utilization of this assay for MTBC/NTM detection and differentiation can avoid the need to wait for culture results from smear-positive samples. This is an important benefit for case management and contact investigation, making the assay especially valuable for clinicians in countries with a high incidence of NTM [23,24]. SO-DMT is effective to identify for *Mycobacterium tuberculosis* complex and differentiation from genus *mycobacterium* in smear positive sputum samples. In this study, SO-DMT is not effective for diagnosis in smear negative sputum samples of MTBC. Wide range prospective study must be designed in smear negative and smear positive sputum samples colobration with clinical data.

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