



Biodiversity and Biogeography of Three *Pseudomonas syringae* Pathovars which Affect Kiwi Fruit Cultivation



Morán FE^{1*}, Marco-Noales E¹, Escrich A², Barbé S¹ and López MM¹

¹ Department of plant protection and biotechnology, Valencian Institute of Agricultural Research (IVIA) Moncada Valencia, Spain

² Universitat de Valencia (UV), Burjassot Valencia, Spain

*Corresponding author: Spain, Email: moran_fel@gva.es

Submission: 📅 September 10, 2018; Published: 📅 November 15, 2018

Abstract

Kiwi fruit is an economically important crop in several countries and it can be affected by several bacterial pathogens. Three pathovars of *Pseudomonas syringae* have been reported affecting kiwi fruit plants: *Pseudomonas syringae* pv. *actinidiae* causal agent of bacterial canker of kiwi fruit; *P. syringae* pv. *actinidifoliorum*, causal agent of leaf spot of kiwi fruit, and *P. syringae*, pv. *syringae* causal agent of floral buds necrosis. Since there is a high phenotypic and genetic diversity among strains inside these pathovars here are briefly described the main differences found in the kiwi fruit strains of these three pathovars to help in developing preventive and curative control strategies to efficiently manage the bacterial diseases that affect kiwi fruit.

Keywords: Biovar; Populations; Phenotypic diversity; Genetic diversity

Introduction

The kiwi fruit is a climbing plant of the genus *Actinidia*, family *Actinidiaceae*. Its cultivation is economically important in many countries being the main world producers China, Italy and New Zealand but in many European and South American countries it is currently considered an emerging crop. According to the latest data available 4.274.870 tons of kiwi fruit were produced in 2016 (<http://www.fao.org/>). The main cultivated species are *A. deliciosa* and *A. chinensis* due to its high fruit production and quality. This emerging crop like many others is not exempt from diseases caused by phytopathogenic agents such as fungi and bacteria.

Among bacteria, three pathovars of the plant pathogen *Pseudomonas syringae* have been reported affecting kiwi fruit plants. Infact the most important disease of kiwi fruit is bacterial canker caused by *P. syringae* pv. *actinidiae* (Psa), which is a limiting factor for the cultivation of susceptible cultivars and has caused important economical losses around the world since 2008. The bacterium was reported for the first time in 1989 in Japan. [1] and then in China [2], Korea and Italy [3,4]. From 2008 onward more aggressive strains of Psa were detected in several countries like Italy [5], New Zealand [6], France [7], and Australia [8] affecting especially some golden yellow fleshed cultivars of *A. chinensis*. Psa is currently considered the most threatening pathogen of some *Actinidia* crops worldwide.

Therefore, it was included in the list of A2 quarantine organisms of the European and Mediterranean Plant Protection Organization

(EPPO) and was the topic of a decision of the european commission (2012) to prevent its introduction in new areas and its dissemination in the European Union. The characteristic symptoms of this disease usually appear in early spring: angular brown spots surrounded by a yellow halo in the leaves and in the branches moreover if the environmental conditions are appropriate trunk cankers with white to reddish exudates can also appear. These symptoms are usually accompanied by wilting and in very susceptible cultivars like 'Hort 16A' even plant death can occur.

There are different populations or biovars of Psa distributed around the world. The characterization and classification in biovars has been designed according to the phenotypic and genetic characterization of the strains from different areas. The principal phenotypic tests used for the characterization of Psa populations are: fluorescence on King's medium B, LOPAT tests (levan production oxidase potato rot arginine dihydrolase and tobacco hypersensitivity), ice nucleating activity (INA), GATTA tests (gelatin liquefaction, aesculin hydrolysis, tyrosinase activity and tartrate utilization) and plant susceptibility. And the principal genetic tests for the characterization are based on the multilocus sequence analysis (MLSA), the type III secretion system effector genes, the repetitive genome sequences (rep-PCR) and the analyses of complete genomes [9,10]

Until 2014 four Psa biovars were described biovar 1 (Psa1) strains were isolated in Japan and Italy before 2008 and were characterized by their low virulence in kiwi plants biovar 2 (Psa2) was only reported in Korea and biovar 3 (Psa3) responsible for

important damages in kiwi crops worldwide is characterized by highly virulent strains [6]. Different studies of the complete genome of several strains show that Chinese strains are the origin of this pandemic lineage [11].

Strains of Psa3 from different countries have been deeply studied at the genetic and phenotypic levels and recently certain differences among strains of this biovar 3 were observed. In Portugal by morphological, biochemical, physiological and molecular tests two genetically different subpopulations of Psa3 have been identified [12]. Moreover, an analysis of New Zealand and European strains of Psa3 using single molecule real-time (SMRT) showed molecular evidences of frequent mobilization and loss of the transposon Tn6212 (without any change in the phenotype) as well as large chromosome inversions and ectopic integration of IS sequences that affect genes *hrp* and is associated with a negative hypersensitivity response (HR) [8]. Other studies have reported that some Italian strains of Psa3 also isolated from symptomatic plants show negative HR [13]. All these evidences suggest a coexistence of different strains and the existence of a gene exchange among different Psa populations from the same source. An analysis of 80 sequenced Psa genomes revealed a very high diversity even within the single clade from which the global pandemic arose [9]. However, a robust core genome can be recognized with minimal impact of within-clade recombination. It was observed in the core genome the lack of genes for N-acyl homoserine lactone (AHL) or the typical complete *luxI/R* QS (quorum sensing) system that is absent in all studied strains [4].

Biovar 4 was created for low virulence strains since the affected kiwi fruit plants did not show trunk cankers, the leaf symptomatology was minor and decreasing production and death of plants were not observed. Recently, through a comparative analysis at the phenotypic, genetic and phylogenetic level, strains of biovar 4 had been reclassified and renamed as a new pathovar *P. syringae* pv. *actinidifoliorum* (Psa_f) [14]. Until now, the strains of pathovar *actinidifoliorum* do not have a significant economic impact on kiwifruit production in the countries where it has been reported, that are France [15], Spain [16] and Japan [17]. Psa_f lineages had been reported suggesting that a high variability exists in this group and that each lineage has its own specific characteristics [14] and [19]. Additionally, new strains that present similar profiles (phenotypic and molecular) to Psa_f but are not identical to the French strains were isolated in Spain. These new strains have been called “Psa_f look alike” they produce a symptomatology similar to Psa_f when inoculated in leaves of kiwi cultivar ‘Hayward’ [20]. The current data also suggest a genetic diversity among them.

Two new biovars, 5 and 6 (Psa₅ and Psa₆), have been recently reported in Japan [21]. Psa₅ was isolated from ‘Hort16’ in Saga Prefecture and Psa₆ was isolated from *A. deliciosa* cv. ‘Hayward’ in Nagano Prefecture. The fact that Psa₅ and Psa₆ had been detected in limited areas suggests that these new biovars may be endemic and rather related to Psa₂. Indeed, a recent study suggests that Psa₅ appears to share an ancestor with the Korean Psa-2 strains [9].

Another *P. syringae* pathovar that affects kiwi fruit but to a lesser extent in comparison to Psa is *P. syringae* pv. *syringae* (Pss) [22], a causal agent of floral buds necrosis that can cause remarkable economic damages in different countries when conditions are favourable for this pathovar. The symptomatology includes canker on twigs, necrotic spots on leaves, browning and rots of flowers and buds [23]. Biodiversity of kiwi strains of this pathovar has not been studied in depth but there is information about populations with INA-positive and negative strains in Italy [24]. Consequently, there is a high genetic and phenotypic diversity of *P. syringae* strains that affect kiwi fruit crops. Biodiversity was also reported in other hosts and is typical of this species complex, divided into 50 pathovar variants and distributed into at least seven distinct phylogroups based on the nucleotide sequence divergence of a small set of housekeeping genes [25, 26, 27].

The *P. syringae* species complex possesses a remarkably broad host range, although individual lineages often show a high degree of host-specificity and with some exceptions as pv. *syringae* re generally constricted to a small number of potential host species. It is also important to remark that inside this complex several processes of gene dynamics and *phytoadaptation* exist, which is a consequence of molecular interactions (by recombination and/or mutations) between different populations. An example of these processes is the mechanism that allows many *P. syringae* pathovars to evade the Effector-Triggered Immunity (ETI) response [28]. In the case of *P. syringae* strains from kiwi, the study of its variability would make it possible to know the role of interactions within the Psa populations and with the native microbiota, which would ultimately help in developing preventive and curative control strategies to efficiently manage the bacterial diseases that affect kiwi fruit.

Acknowledgement

The authors thank Monterde A, Penyalver J, González A and Landeras E for the excellent technical support and F. X. Silva-Hernandez for the English edition. The IVIA group acknowledges the grants from INIA for RTA 2013-00072-C03-O3 and the support of Spanish Ministry of Agriculture to the National Reference of Laboratory for Phytopathogenic Bacteria.

References

1. Takikawa Y, Serizawa S, Ichikawa T, Tsuyumu S, Goto M (1989) *Pseudomonas syringae* pv. *actinidiae* pv. *nov.* The causal bacterium of canker of kiwifruit in Japan. *Japanese Journal of Phytopathology* 55(4): 437-444.
2. Wang Z, Tang X, Liu S (1992) Identification of the pathogenic bacterium for bacterial canker on Actinidia in Sichuan. *Journal of Southwest Agricultural University, China*.
3. Koh JK, Cha BJ, Chung HJ, Lee DH (1994) Outbreak and spread of bacterial canker in kiwifruit. *Korean Journal of Plant Pathology* 10(1): 68-72.
4. Scortichini M (1994) Occurrence of *Pseudomonas syringae* pv. *actinidiae* on kiwifruit in Italy. *Plant Pathology* 43(6): 1035-1038.
5. Patel HK, Ferrante P, Covaceuszach S, Lamba D, Scortichini M et al. (2014) The kiwifruit emerging pathogen *Pseudomonas syringae* pv.

- actinidiae* does not produce AHLs but possesses three luxR solos. PLoS One 9(1): e87862.
6. Everett KR, Taylor RK, Romberg MK, Rees GJ, Fullerton RA, et al. (2011) First report of *Pseudomonas syringae* pv. *actinidiae* causing kiwifruit bacterial canker in New Zealand. Australasian Plant Disease Notes 6(1): 67-71.
 7. Vanneste JL, Poliakov F, Audusseau C, Cornish DA, Paillard S, et al. (2011) First report of *Pseudomonas syringae* pv. *actinidiae*, the causal agent of bacterial canker of kiwifruit in France. Plant Disease 95(10): 1311-1311.
 8. EPPO (2011) First report of *Pseudomonas syringae* pv. *Actinidiae* in Australia, Australia.
 9. Firrao G, Torelli E, Polano C, Ferrante P, Ferrini F, et al. (2018) Genomic structural variations affecting virulence during clonal expansion of *Pseudomonas syringae* pv. *actinidiae* biovar 3 in Europe. Frontiers in Microbiology. Frontiers Media SA 9(5): 656.
 10. McCann HC, Rikkerink EHA, Bertels F, Fiers M, Lu A (2013) Genomic analysis of the kiwifruit pathogen *Pseudomonas syringae* pv. *actinidiae* provides insight into the origins of an emergent plant disease. PLoS Pathogens 9(9): e1003503.
 11. McCann HC, Li L, Liu Y, Li D, Pan H (2017) Origin and evolution of the kiwifruit canker pandemic. Genome Biology and Evolution 9(4): 932-944.
 12. García E, Moura L, Abelleira A, Aguin O, Ares A, et al. (2018) Characterization of *Pseudomonas syringae* pv. *actinidiae* biovar 3 on kiwifruit in north-west Portugal. J Appl Microbiol 125(4): 1147-1161.
 13. Biondi E, Zamorano A, Vega E, Ardizzi S, Sitta D, et al. (2018) Draft whole genome sequence analyses on *Pseudomonas syringae* pv. *actinidiae* hypersensitive response negative strains detected from kiwifruit bleeding sap samples. Phytopathology 108(5): 552-560.
 14. Cuntly A, Poliakov F, Rivoal C, Cesbron S, Fischer Le Saux M et al. (2014) Characterization of *Pseudomonas syringae* pv. *actinidiae* (Psa) isolated from France and assignment of Psa biovar 4 to a de novo pathovar: *Pseudomonas syringae* pv. *actinidifoliorum* pv. nov. Plant Pathology 64(3): 582-596.
 15. Berge O, Monteil CL, Bartoli C, Chandeysson C, Guilbaud C, et al. (2014) A user's guide to a data base of the diversity of *Pseudomonas syringae* and its application to classifying strains in this phylogenetic complex. PLoS One 9(9): e105547.
 16. Abelleira A, Ares A, Aguin O, Picoaga, A, Lopez MM, et al. (2015) Detection and characterization of *Pseudomonas syringae* pv. *actinidifoliorum* in kiwifruit in Spain. Journal of Applied Microbiology 119(6): 1659-1671.
 17. Sawada H, Fujikawa T, Kita N, Orihara N, Shinozakiz T (2017) Characteristics of *Pseudomonas syringae* pv. *actinidifoliorum* causing bacterial leaf spot of *Actinidia* spp. in Japan. Japanese Journal of Phytopathology 83(3): 136-150.
 18. Sawada H, Kondo K, Nakaune R (2016) Novel biovar (biovar 6) of *Pseudomonas syringae* pv. *actinidiae* causing bacterial canker of kiwifruit (*Actinidia deliciosa*) in Japan. Japanese Journal of Phytopathology 82(2): 101-115.
 19. Morán F, Landeras E, Peñalver J, Monterde A, Morente C (2016) Libro de resúmenes del XVIII Congreso de la Sociedad Española de Fitopatología. PL-103 Caracterización de nuevas cepas de *Pseudomonas syringae* aisladas de kiwi y con baja virulencia. Palencia, p. 77.
 20. Sawada H, Miyoshi T, Ide Y (2014) Novel MLSA group (Psa5) of *Pseudomonas syringae* pv. *actinidiae* causing bacterial canker of kiwifruit (*Actinidia chinensis*) in Japan. Japanese Journal of Phytopathology 80(3): 171-184.
 21. Balestra GM, Varvaro L (1997) *Pseudomonas syringae* pv. *syringae* causal agent of disease on floral buds of *Actinidia deliciosa* (A. Chev) Liang et Ferguson in Italy. Journal of Phytopathology 145(8-9): 375-378.
 22. Gagnard J, Luisetti J (1992) Role du pouvoir glucogène dans le processus infectieux de *Pseudomonas syringae* pv. *syringae* et de *Pseudomonas viridiflava* sur kiwi. Fruits 47(4): 495-501.
 23. Rossetti A, Balestra GM (2008) *Pseudomonas syringae* pv. *syringae* on kiwifruit plants: Epidemiological traits and its control. *Pseudomonas syringae* pathovars and related pathogens- Identification, epidemiology and genomics. Springer Dordrecht.
 24. Hwang MSH, Morgan RL, Sarkar SF, Wang PW, Guttman DS (2005) Phylogenetic characterization of virulence and resistance phenotypes of *Pseudomonas syringae*. Applied and Environmental Microbiology 71(9): 5182-5191.
 25. Sarkar SF, Guttman DSN (2004) Evolution of the core genome of *Pseudomonas syringae*, a highly clonal, endemic plant pathogen. Applied and Environmental Microbiology 70(4): 1999-2012.
 26. Parkinson N, Bryant R, Bew J, Elphinstone J (2010) Rapid phylogenetic identification of members of the *Pseudomonas syringae* species complex using the *rpoD* locus. Plant Pathology 60(2): 338-344.
 27. Lindeberg M, Cunnac S, Collmer A (2009) The evolution of *Pseudomonas syringae* host specificity and type III effector repertoires. Molecular Plant Pathology 10(1): 767-775.



Creative Commons Attribution 4.0 International License

For possible submissions Click Here

[Submit Article](#)



Biodiversity Online Journal

Benefits of Publishing with us

- High-level peer review and editorial services
- Freely accessible online immediately upon publication
- Authors retain the copyright to their work
- Licensing it under a Creative Commons license
- Visibility through different online platforms