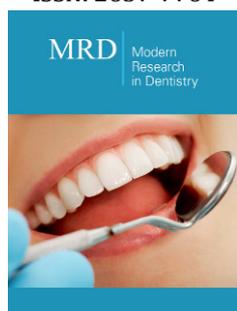


Contour Augmentation after Guided Bone Regeneration with Porous Titanium Granules: A Clinical, Histologic and Histomorphometric Evaluation

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Abstracts

Objectives: To determine clinical, histological, and histomorphometric results following ridge preservation via contour augmentation with high resistance biomaterials.

Material and Methods: A 54-year-old female was referred by her general dentist due to her crowded and periodontally hopeless maxillary right canine and premolar teeth. Two dental implants were surgically placed in palatal positions. The exposed implant surfaces were covered with a slowly-resorbable, synthetic, nanocrystalline hydroxyapatite bone substitute and overbuilding was done with porous titanium granules (PTGs) mixed with the patient's blood. A long-lasting collagen membrane was fixed over the graft sites. Patient was recalled six months later for the second-phase surgery. At that time, a core biopsy from the augmentation site was taken.

Result: The patient's cone beam computed tomography scan showed that hard tissue width and height increased from 8.4 and 10.6mm to 9.2 and 12.3mm, respectively. The histological sections revealed that new bone was bridging between the PTGs and neighboring particles. The new bone matrix consisted of 12.41% mineralized bone matrix and 2.82% osteoid. Concerning the osteoconductivity of PTGs, 27.0% new mineralized bone, 10.2% osteoid, and 62.8% soft tissue were found covering the titanium particles.

Conclusion: Overbuilding the ridge via contour augmentation with non-resorbable titanium granules could serve as a valid approach based on optimal clinical and biological results.

Clinical Relevance: Immediate implant placement and use of PTGs and collagen membrane all at the same time can improve clinical periodontal parameters and implant stability in the short-term.

Keywords: Guided Bone Regeneration; Contour Augmentation; Ridge preservation; Titanium Granules

Introduction

Tooth extraction causes inevitable changes in the supporting structures, which may lead to complications for dental implant placement [1]. Since the 1960s, many studies have been conducted to assess the loss of hard and soft tissues after tooth extraction, and innumerable efforts have been made to prevent or minimize bone loss [2]. Different methods have been suggested to preserve and reconstruct bone volume and prevent alveolar ridge resorption following traumatic events such as tooth extraction. The guided bone regeneration (GBR) technique has presented promising results in repairing bone defects. Autogenous bone grafts are considered the gold standard for GBR, due to their osteoconductive, osteoinductive, and osteogenic properties [3,4]. Despite their high efficacy for bone reconstruction, the need for

a second surgical site, unpredictable remodeling rate and bone loss have been regarded as their main disadvantages.

Thus, more recent studies have aimed to find a suitable alternative for autogenous bone grafts. Bone substitutes have found their niche within the field of dentistry and have shown promising results [5-7]. Hydroxyapatite-based materials are commonly used for this purpose. Rothamel et al. [8] evaluated the efficacy of nanocrystalline hydroxyapatite paste; however, he claimed that it is not efficient for ridge preservation due to its unpredictable resorption pattern. Since nonresorbable materials can withstand external loads and are resistant to deformation, they can be used with high success rate in bone defect reconstruction, especially for contour augmentation [5, 7,9-11]. The biocompatibility of titanium has been proven in the recent years, and its use in implant dentistry and orthopedics is widely growing. Titanium is highly resistant to corrosion in body fluids. Furthermore, it is considered a potentially appropriate bone substitute material due to its nonresorbable properties [10,11].

Titanium particles can stimulate the activation of complement system and platelets and can consequently increase the level of platelet-derived growth factor (PDGF). PDGF has been shown to promote bone growth, and this capability along with large surface area is advantageous for bone reconstruction [12]. Porous titanium granules (PTG) (Natix™, Tigran Technologies AB, Malmo, Sweden) possess these properties and contain 700-1000µm diameter granules. The porous nature of these granules enables bone infiltration between the particles. PTGs consist of irregular, highly porous, nonresorbable granules of commercially pure titanium; they were first introduced for treatment of peri-implant defects. When implanted, the granules are able to interlock with each other, providing a suitable environment for intra- and inter-granule osteogenesis. A stable porous structure is formed as such, which provides an optimal environment for bone ingrowth. Moreover, PTGs have been successfully used in different circumstances within the specialty of implant dentistry. They have also been suggested for management of periodontal furcation involvement of teeth [13-15]. On the basis of the mean total amount of regenerated bone, Tavakoli et al. [16] reported that the use of Natix™ bone substitute with a membrane can promote bone regeneration in the process of healing of extraction sockets in dogs [16]. Considering the valuable characteristics properties of titanium granules as nonresorbable materials, they may be suitable for contour augmentation with different GBR procedures. Therefore, the purpose of this case report was to determine post-extraction dimensional changes following ridge preservation via contour augmentation with PTGs and a collagen membrane for horizontal GBR. As a second objective, the histological composition of the grafted extraction socket was evaluated after 8 months of follow-up.

Material and Methods

Patient

A 54-year-old female was referred to our clinic complaining of crowded and periodontally hopeless maxillary right canine and premolar teeth. Due to excessive loss of tooth structure and severe

bone loss detected circumferentially, the suggested treatment plan included extraction of teeth (right canine and premolars) and immediate implantation with GBR. Initial cone beam computed tomography data showed that there was no buccal bone plate over the existing teeth (Figure 1).

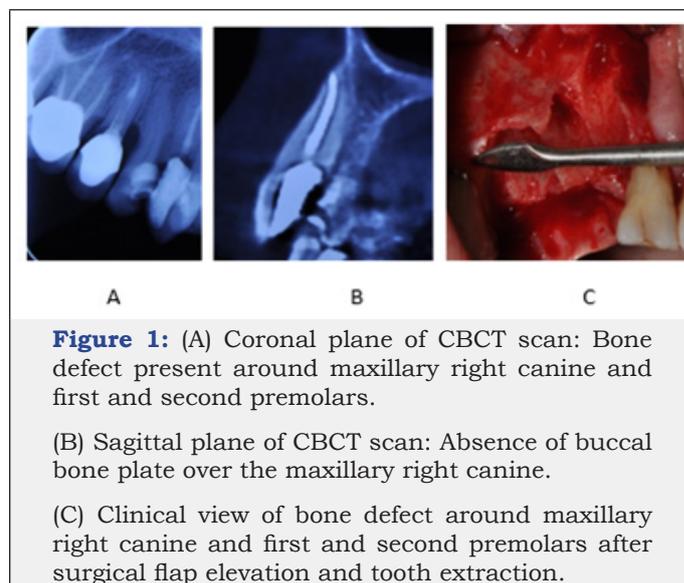


Figure 1: (A) Coronal plane of CBCT scan: Bone defect present around maxillary right canine and first and second premolars.

(B) Sagittal plane of CBCT scan: Absence of buccal bone plate over the maxillary right canine.

(C) Clinical view of bone defect around maxillary right canine and first and second premolars after surgical flap elevation and tooth extraction.

Presurgical treatment

The patient's medical and dental history showed that she had no contraindication for dental implant surgery. The cone beam computed tomography scans, periapical radiographs and clinical photographs were obtained, dental cast was poured, and a clinical periodontal examination was carried out, which included assessment of bleeding on probing (BOP), pocket depth, and plaque index. The patient underwent scaling and root planning and oral hygiene and plaque control instructions were reinforced.

Surgical treatment (stage 1)

Local anesthesia was administered at the surgical site (buccally and palatally) using 2% lidocaine with 1:100,000 epinephrine. A sulcular incision was made extending from tooth #2 to tooth #6 with a releasing incision at the mesial of tooth #6. A full thickness mucoperiosteal flap was elevated to expose the labial and palatal bone plate. Interdental papillae were preserved and reflected buccally. The hopeless teeth (canine and premolars) were extracted via a conservative approach using piezo surgery and periostomes. The tooth extraction sockets were then curetted to remove all soft tissue remnants. The bone defects and surgical sites were thoroughly irrigated with sterile saline solution. Two dental implants (4.2x11.5mm SIC max for anterior socket and 4.0x11.5mm SIC ace for posterior socket; SIC, Switzerland) were installed in palatal positions. Due to inadequate buccal bone, 4-8 threads of inserted implants remained exposed. The exposed implant surfaces were covered with a slowly-resorbable, synthetic, nanocrystalline hydroxyapatite bone substitute (NanoBone®, Artoss, Germany) and overbuilding was done beyond the resorbed or lost buccal plate with PTGs (Natix™, Tigran Technologies AB, Malmo, Sweden) mixed with the patient's blood (obtained from a vein). A long-lasting collagen membrane (SIC membrane; SIC, Germany) was

fixed with two tacks (FRIOS bone fixation system, FRIOS, Germany) at the most apical parts in the buccal side and subsequently at the palatal side to firmly cover the entire grafted area. The membrane covered not only the defect site, but also a 3mm margin of sound bone at the mesial, distal, apical and palatal.

The flaps were closed with tension-free sutures (3-0 silk) (Figure 2). Postoperatively, the patient was instructed to take 500mg amoxicillin three times a day for a total of 10 days. Also, 0.2% chlorhexidine gluconate mouth rinse (DarouPakhsh Holding Co.,

Iran) was prescribed for plaque control, twice daily. The patient's initial postoperative visit was 10 days after surgery; the silk sutures were removed. The patient received professional cleaning on a daily basis alternately between a periodontist and general dentist for the first two weeks. Mechanical oral hygiene practice, consisting of tooth brushing and inter-dental brushing, was initiated by the patient on the 28th post-operative day. Post-operative care included oral hygiene reinforcement and professional plaque removal done on demand.

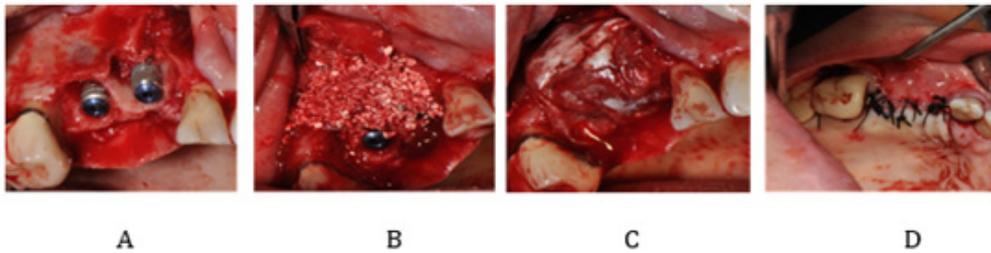


Figure 2: (A) Placement of two implants at the sites of maxillary right canine and second premolar teeth. (B) Covering the buccal bone defect with PTGs. (C) Placement of collagen membrane over PTGs. (D) Returning the flap and suturing.

Surgical treatment (stage 2)

The patient was recalled six months later for the second-phase surgery and uncovering of implants. A 2.7mm x 6.0mm trephine bur was used to take a core biopsy from the augmentation site under copious saline irrigation. The biopsy core was immersed in

10% formalin solution. The patient was followed-up weekly until completion of soft tissue healing and then monthly until healing abutment placement (Figure 3). The trephine biopsy core was sent to the Robert K. Schenk Laboratory of Oral Histology (School of Dental Medicine, University of Bern, Switzerland) for histologic and histometric evaluation.

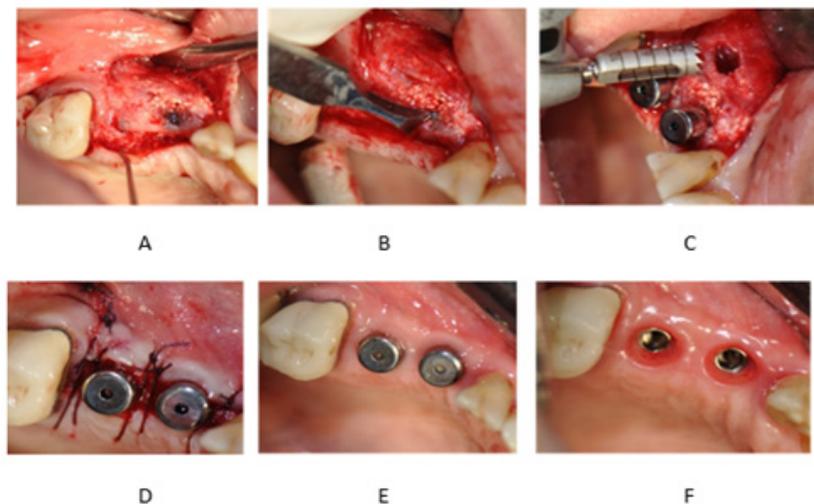


Figure 3: (A) Flap elevation in the second-stage surgery. (B) Uncovering the implants. (C) Obtaining a core biopsy. (D) Returning the flap and suturing. (E) Soft tissue healing six months after surgery. (F) Opening of healing abutment to place the temporary abutment.

Sample preparation and descriptive histology

The specimen was dehydrated in ethanol series and embedded in methyl methacrylate. Undecalcified sections measuring ~500 microns in thickness were cut using a low speed diamond saw under coolant (Varicut® VC-50, Leco, Munich, Germany). After mounting the sections on acrylic glass slabs, they were ground and polished to a final thickness of about 100µm (Knuth-Rotor-3, Struers, Rodovre/Copenhagen, Denmark). Most sections were stained with toluidine blue and basic fuchsin and the two most central ground sections were used for qualitative and quantitative analyses. A few sections were used for polarized light microscopy. Digital photography was performed using a ProgRes® C5 digital camera (Jenoptik Laser, Optik Systeme GmbH, Jena, Germany) connected to a Zeiss Axioplan microscope (Carl Zeiss, Göttingen, Germany). Image processing and morphometric analyses were performed directly under the microscope. One experienced examiner (D.B.) performed descriptive histological analysis.

Histomorphometry

The area fractions (%) of the newly formed mineralized bone, osteoid, filler material 1 (PTG), filler material 2 (NanoBone) and soft tissue (ST) occupying the biopsy volume were determined by point counting directly under the microscope, using a square grid (distance between 6x6 test lines: 0.067mm) at x250 magnification. The percentage of PTG surface covered with the newly formed bone (osteoconduction) was evaluated by counting the intersections using the same grid and magnification as mentioned above.

Result

Clinical results

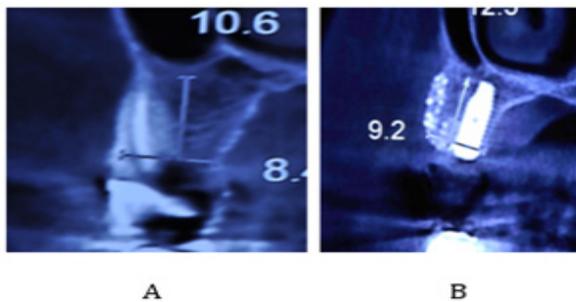


Figure 4: (A) Assessment of bone width and height on preoperative CBCT scan of patient;

(B) Assessment of bone width and height on postoperative CBCT scan of patient.

Our patient was a 55-year-old female with no systemic disease, who was referred to our clinic due to failed endodontic treatment of the maxillary right canine and premolar teeth. After extraction of the afore-mentioned teeth due to their hopeless prognosis, two implants were surgically placed at the sites of the maxillary right canine and second premolar. After implant surgery, hard and soft tissue assessment was meticulously done until prosthetic loading of implants six months after the second-stage surgery every two months. During the follow-up sessions, parallel periapical radiographs were obtained, scaling and root planning

was performed, and the oral hygiene instructions were given and reinforced. Periodontal parameters including pocket depth, BOP and pus formation were also assessed. Clinical assessment showed no signs of inflammation around the dental implants. Probing depth in the buccal, palatal and interproximal areas of implants was 6 mm and BOP was present. This value was 3mm without BOP or pus after treatment. The patient's cone beam computed tomography scan showed that width and height of bone increased from 8.4mm and 10.6mm before treatment to 9.2mm and 12.3mm after treatment, respectively (Figure 4). No signs of graft loss or gingival recession were noted around dental implants.

Histology

The histological sections revealed the presence of PTGs, new bone, soft tissue and NanoBone® (Figure 5-7). The PTGs were variable in shape, size and density. Most of the new bone was lamellar bone (Figure 6a); however, osteoid lined by osteoblasts was occasionally observed (Figure 6b). The intervening soft tissue was non-fibrous and rich in small and medium-sized blood vessels (Figure 6a,c). New bone was observed in direct contact with PTGs (Figure 6c) and in the pores of PTGs (Figure 6d). Furthermore, new bone was bridging between PTGs and neighboring NanoBone® particles (Figure 7a). The NanoBone® particles were lined by new bone (Figure 7a) and few multinucleated, osteoclast-like, giant cells (Figure 7b). Such multinucleated giant cells were not observed on the PTG surface. The soft tissue peripheral to the NanoBone® particles was cell-rich (Figure 7b).

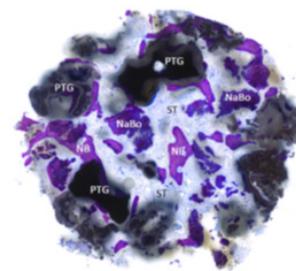


Figure 5: Histological section revealed the presence of PTG: porous titanium granules, NB: new bone, ST: soft tissue, and NaBo: remnant particles of nano hydroxyapatite synthetic bone substitutes.

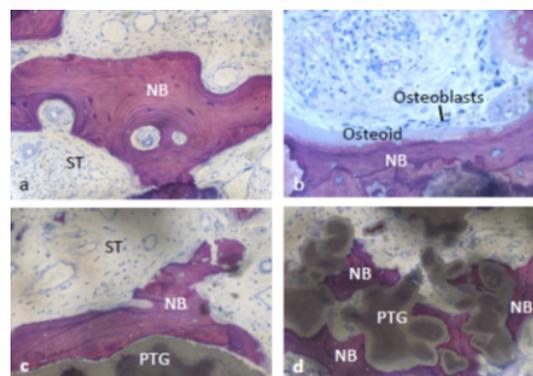


Figure 6: ST: soft tissue and NB: New bone in direct contact with PTGs: porous titanium granules.

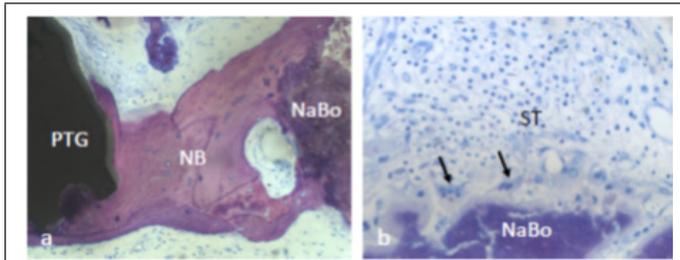


Figure 7: NB: new bone was bridging between PTGs: porous titanium granules and neighboring NaBo: synthetic nano hydroxyl apatite bone substitutes (a). Multinucleated, osteoclast-like, giant cells (arrows) and cell-rich soft tissue were observed peripheral to Nano Bone particles (b).

Histomorphometry

The area fractions of new bone, soft tissue, PTGs, and NanoBone[®] were 15.2%, 39.3%, 22.4%, and 23.1%, respectively (Figure 8a). The new bone matrix consisted of 12.41% mineralized bone matrix and 2.82% osteoid. Concerning the osteoconductivity of PTGs, 27.0% new mineralized bone, 10.2% osteoid, and 62.8% soft tissue were found to cover the titanium particles (Figure 8b).

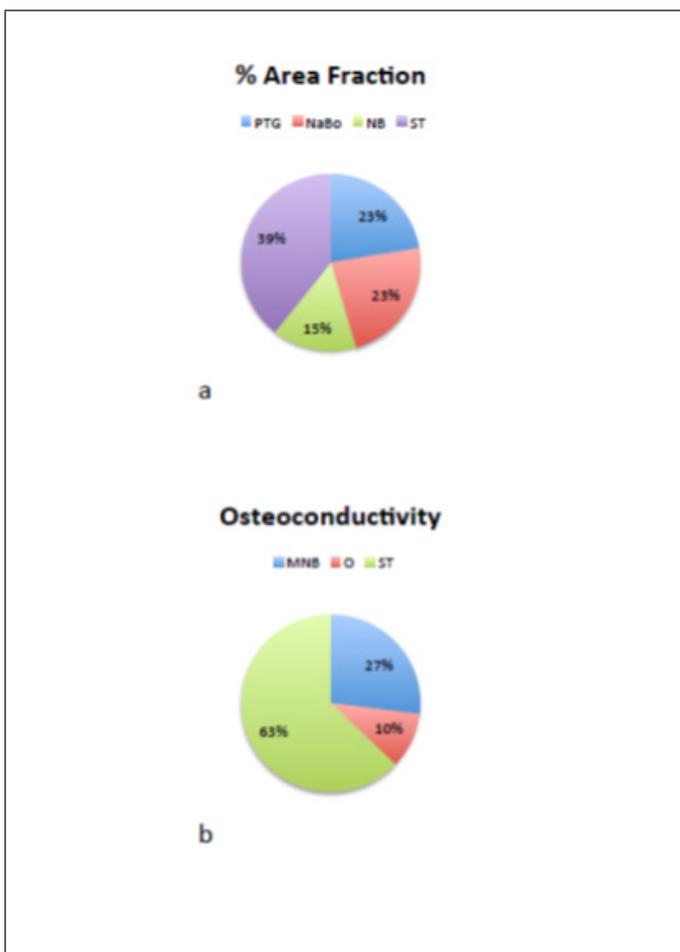


Figure 8: The area fractions of NB: new bone, ST: soft tissue, PTGs: porous titanium granules, and NaBo: NanoBone (a). The new bone matrix consisted of MNB: mineralized bone matrix, ST: soft tissue, and O: osteoid (b).

Discussion

In this case study, PTGs were used to cover the bone defect in the buccal surface of implants in the first phase of surgery. PTGs were first introduced by Bystedt et al. [17] as an osteoconductive material for bone grafting in sinus floor augmentation surgery [17]. Today, PTGs are used for treatment of bone defects [18], inflammatory peri-implant defects [19] and healing of tooth extraction sockets [20]. In our study, some clinical periodontal parameters such as pocket depth, BOP and pus formation were assessed at baseline and every two months after the second-stage surgery. Wohlfahrt et al. [21] reported that clinical attachment level and gingival recession were not significantly different from the baseline values 12 months after treatment of grade II furcal defects of mandibular molars with PTG, but pocket depth and gingival index significantly decreased after treatment and BOP remained unchanged [14]. An animal study by Wohlfahrt et al. [21] showed that probing depth increased six weeks after treatment of grade II furcal defects with PTGs compared to baseline, but gingival index decreased after treatment. Horizontal probing depth was lower in group treated with PTGs compared to the deproteinized bovine bone mineral and sham groups, although the difference between the groups was not significant [21].

In our study, pocket depth was 6mm before treatment and BOP was present in the areas with bone loss around teeth. After implant placement and bone grafting, pocket depth improved to 3mm and BOP was no longer present. For bone grafting and dental implant placement, graft material may be applied before or simultaneously with placement of dental implants at the site of bone loss. Applying bone graft material prior to implant placement requires an additional surgical procedure, and the associated risks and costs must be taken into account. In our study, bone grafting and dental implant placement were done simultaneously. Table 1 [22-34] lists some studies that used this technique with different bone graft materials. In all the studies listed in Table 1 [22-34], this technique yielded favorable radiographic and clinical results. This technique eliminates the need for an additional surgical procedure; however, there is a risk of implant movement and subsequent movement of graft material. Therefore, this technique requires high precision, and immediate loading of implants is not encouraged and should be avoided.

Moreover, stable bone graft materials must be used. In our study, implant loading was performed six months after the second-stage surgery and a resorbable membrane was placed over PTGs for further stabilization. In most of the studies listed in Table 1 [22-34], GBR technique was used for bone grafting. The results of a review study by Aghaloo et al. [35] showed that the implant survival rate in GBR technique for augmentation of edentulous maxillary ridge was between 96.1-100%, which was slightly higher than that of other technique [36]. Also, collagen membrane was used in the majority of the studies cited in Table 1 and also in our study. Collagen membrane has excellent biocompatibility [37] and more flexibility than ePTFE and titanium mesh [38] and does not need additional surgery for its removal. It appears that use of membrane along with regenerative material enhances the short-

term results of bone augmentation since Delgado Ruiz et al. [39] showed that six weeks after bone defect coverage using PTGs along with collagen membrane, the percentage of cortical defect closure was significantly higher than that of the PTG group without the collagen membrane. In the PTG group without collagen membrane, evidence of tissue inflammation was noted [40]. Roos et al. [41] stated that use of resorbable membrane along with bone substitute

had no significant difference from the use of bone substitute alone for treatment of peri-implantitis defects with regard to the improvement of clinical and radiographic parameters in long-term (5 years) [42]. The above-mentioned findings indicate that as long as the membrane has not been resorbed, it protects bone graft materials against movement and degradation and can therefore enhance the short-term results [43,44].

Table 1: Studies on simultaneous placement of implant and different bone graft material.

	Author/Year	Study Groups	Graft Material	Immediate or Delayed Implantation	Implant Placement Surgical Technique	Human/Animal Study	Study Design
1	Janner, et al. [36]	1- A+D+M 2- A+D 3- D+M 4- D	A: autogenous bone chips, D: DBBM, M: collagen membrane	Delayed	-	Animal	preclinical study
Result: Addition of autogenous bone chips and presence of collagen membrane increased bone formation around DBBM particles.							
2	Wessing, et al. [37]	1- CXP 2- Reference membrane (BG)	Creos Xenoprotect reference membrane	-	Two-stage	Human	RCT
Result: In the CXP arm, the mean defect height at implant insertion site was 5.1±2.1mm (n=24), which decreased at reentry by 81% to 1.0±1.3mm (n=23). In the BG arm, the defect height at implant insertion site was 4.9±1.9mm (n=25) and reduced at reentry by 62% to 1.7±2.1mm (n=24).							
3	Benic, et al. [38]	1- GBR: DBBM+AB+CM 2-Standard implant placement without GBR	DBBM, autogenous bone, collagen membrane	-	-	Human	15-year results
Result: Implants placed with simultaneous GBR by using particulate DBBM and/or AB in combination with CM did not significantly differ from implants completely placed into pristine bone with respect to 15-year implant survival, interproximal bone levels, and dimensions of buccal bone and mucosa.							
4	Benic, et al. [40]	1- Without GBR 2- With GBR	DBBM, collagen membrane	Delayed	-	Human	Prospective study
Result: Implant placement with simultaneous GBR resulted in more gain of buccal soft tissue contour in comparison with implant placement without GBR.							
5	Gultekin, et al. [42]	1- Autogenous iliac bone grafting 2- Nonresorbable membrane	Autogenous iliac bone grafting, nonresorbable membrane	-	-	Human	Retrospective study
Result: Although both hard tissue augmentation approaches provide an adequate volume of bone graft for implant insertion, IBG results in greater graft resorption at maxillary augmented sites than GBR.							
6	Demarosi, et al. [22]	Regenerative materials and a barrier	regenerative materials and a barrier	Immediate	-	Human	A Technical Note
Result: This procedure preserves the apical ridge bone crest, allowing implant anchorage and primary stability to be achieved.							
7	Stricker, et al. [23]	1-Mucoperiosteal flap+GBR 2-Mucosal flap	Biphasic calcium phosphate, collagen membrane	-	-	Animal	Preclinical study
8	Kolerman, et al. [24]	Mineralized allograft (FDBA), collagen membrane	Mineralized allograft (FDBA), collagen membrane	Immediate	One-stage	Human	Retrospective study
Result: During the follow up period, the study group presented a mean mesial bone loss of 1.10±0.39mm, and mean distal bone loss was 1.19±0.41mm. Anterior maxillary single-tooth replacement, using GBR and connective tissue graft according to the concept of immediate implant placement, and non-functional restoration is an accepted treatment modality achieving favorable peri-implant soft tissue condition.							
9	Jung, et al. [25]	1- Xenogeneic bone mineral+ CM 2- Xenogeneic bone mineral+ PEG	Xenogeneic bone mineral, collagen membrane, polyethylene glycol membrane	-	-	Human	RCT
Result: A polyethylene glycol membrane used for bone regeneration of peri-implant defects performed as successfully as a collagen membrane with respect to implant survival rate and dimensions of the buccal peri-implant bone and mucosa after 5 years.							
10	Konstantinidis, et al. [26]	1- CPS+CM 2- CPS+TM	Calcium Phosphosilicate, alloplastic bone putty, collagen membrane, titanium mesh	Delayed	Two-stage	Human	Case series
Result: Mean gain in bone height was 3.23±2.04mm, with 75 % of the peri-implant defects achieving complete regeneration. The handling characteristics of CPS putty may simplify GBR protocol. Implants placed in conjunction with GBR have a very good survival rate after one year of follow-up.							

11	Covani, et al. [27]	Porcine bone, collagen membrane	Porcine bone, collagen membrane	Immediate	-	Human	Single-cohort study
Result: The data of this study suggested that the placement of a dental implant by means of a flapless technique in a fresh extraction socket filled with slowly resorbable graft biomaterial and with a delayed prosthetic restoration shows positive final esthetic outcomes. At the 5-year point of the survey, the changes in the bone level were minimal. Moreover, both midfacial tissue and the papillae maintained the early increase recorded before the first year of the follow-up.							
12	Noelken, et al. [28]	Autogenous bone chips	Autogenous bone chips	Immediate	-	Human	2- year results
Result: Even implant sites with facial bone defects can be successfully treated with a favorable esthetic outcome using the immediate implant insertion, immediate reconstruction, and immediate provisionalization technique.							
13	Cucchi, et al. [29]	Heterologous porcine-derived biomaterial, high-density polytetrafluoroethylene	Heterologous porcine-derived biomaterial, high-density polytetrafluoroethylene	Delayed	Two-stage	Human	Case report
Result: the use of a heterologous bone substitute alone to restore a vertical defect in a GBR procedure can be as effective as the standard protocol, while avoiding the drawbacks associated with a second surgical site opening.							
14	Jung, et al. [30]	1- DBBM+CM 2- DBBM+ e-PTFE 3- Control	DBBM, collagen membrane, Expanded polytetrafluoroethylene	-	-	Human	Original study
15	Favero, et al. [31]	1- DBBM+CM 2- Control	DBBM, collagen membrane	Immediate	-	Animal	Experimental study
Result: At the buccal aspect, the most coronal bone-to-implant contact was located at a similar distance from the implant margin at the test (1.7±1.0mm) and control (1.6±0.8mm) sites, respectively. The use of DBBM particles, concomitantly with a collagen membrane, did not additionally improve the outcome obtained at the control sites.							
16	Caneva, et al. [32]	1-DBBM+CM 2-No treatment augmentation	DBBM, collagen membrane	Immediate	-	Animal	Experimental study
Result: The area of the buccal shrinkage of the alveolar crest was significantly smaller at the test sites (5.9±2.4mm ²) compared with the control sites (11.5±1.7mm ²). After 4 months of healing, no differences in soft tissue dimensions were found between the test and control sites based on the histological assessments.							
17	Shibly, et al. [33]	Freeze-dried bone allograft, collagen membrane	Freeze-dried bone allograft, collagen membrane	Immediate	-	Human	Case series
Result: The mean reduction in vertical defects between baseline and re-entry for all sites was 2.42mm (p=0.0005). Compared to lingual sites, the buccal sites showed the greatest resolution in vertical defect dimensions (6.37mm), followed by proximal sites (0.78mm). The overall mean reduction in horizontal defects was 1.59mm (p<0.0001). Compared to lingual sites, the buccal sites showed the greatest resolution in horizontal gap dimension (3.2mm), followed by proximal sites (0.8mm).							
18	Benić, et al. [34]	1- Xenogeneic bone substitute material, autogenous bone or a mixture of the two and defect covering with a bio-absorbable collagen membrane 2- Standard implantation procedure without bone regeneration	Xenogeneic bone substitute material, autogenous bone, bio-absorbable collagen membrane	-	-	Human	5-Year results
Result: Cumulative survival rates reached 100% for the GBR group and 94.1% for the control group without statistical significance. No statistically significant differences for clinical and radiographic parameters were found between the two groups.							

Conclusion

Tooth extraction followed by implant placement and use of PTGs and collagen membrane all at the same time can improve clinical periodontal parameters and implant stability in the short-term. Long-term randomized clinical trials with large sample size are required in this respect. Moreover, future studies are recommended to compare different bone graft materials for simultaneous placement with implant insertion.

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